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1.0 **Scope**

This analytical method specifies general procedures and requirements to follow during crime scene investigation for the collection and preservation of forensic evidence. This manual is applicable to all forensic scientists responding to any type of crime scene investigation.

This analytical method is intended to be followed to ensure the highest level of quality and safety for crime scene investigation.
2.0 **References**

2.1 ACSO Policy & Procedure Manual

2.2 Ada County Forensic Lab Quality Assurance Manual

2.3 Ada County Forensic Lab Health and Safety Manual

2.4 Ada County Forensic Lab Crime Scene Training Manual

2.5 Equipment Maintenance and Operation Manuals
3.0 Terms and Definitions

3.1 **ABFO scales**: (American Board of Forensic Odontology scales). An L-shaped piece of plastic used in photography that is marked with circles, black and white bars, and 18-percent gray bars to assist in distortion compensation and provide exposure determination. For measurement, the plastic piece is marked in millimeters.

3.2 **Adhesive Lifter**: Any of a variety of adhesive coated materials or tapes used for lifting evidence.

3.3 **Alternate light source**: Equipment used to produce visible and invisible light at various wavelengths to enhance or visualize potential items of evidence (fluids, fingerprints, clothing fibers, etc.).

3.4 **Ambient light**: The available or existing light that surrounds the object being photographed.

3.5 **Bindle paper**: Clean paper folded to contain trace evidence, sometimes included as part of the packaging for collecting trace evidence.

3.6 **Biohazard bag**: A container for materials that have been exposed to blood or other biological fluids and have the potential to be contaminated with hepatitis, HIV, or other pathogens.

3.7 **Biological fluids**: Fluids of human or animal origin that are commonly encountered at crime scenes (e.g., blood, mucus, perspiration, saliva, semen, vaginal fluid, urine).

3.8 **Biological weapon**: Biological agents used to threaten human life (e.g., anthrax, smallpox, or any infectious disease).

3.9 **Bloodborne pathogen**: Infectious, disease-causing microorganisms that may be found or transported in biological fluids.

3.10 **Bloodstain Pattern Analysis**: The examination of the size, shape, distribution, and patterns of bloodstains.

3.11 **Body fluids**: Blood, semen, blood products, vaginal secretions, cerebrospinal fluid, synovial fluids, pleural fluid, peritoneal fluid, pericardial fluid, and amniotic fluid. Care should also be taken with other biological materials such as body parts and tissues, saliva, urine, feces.

3.12 **Boundaries**: The perimeter or border surrounding potential physical evidence related to the crime.

3.13 **Case file**: The collection of documents comprising information concerning a particular investigation. (This collection may be kept in case jackets, file folders, ring binders, boxes, file drawers, file cabinets, or digital files. Sub-files are often used within case files to segregate, laboratory requests and reports, evidence documentation, photographs, videotapes, audiotapes, and other documents.)
3.14 **Case identifiers**: The alphabetic and/or numeric characters assigned to identify a particular case.

3.15 **Casting**: The filling of a three-dimensional impression with material that takes on and retains the characteristics which were left in that impression.

3.16 **Chain of custody**: A process used to maintain and document the chronological history of the evidence. (Documents should include name or initials of the individual collecting the evidence, each person or entity subsequently having custody of it, dates the items were collected or transferred, agency and case number, victim’s or suspect’s name, and a brief description of the item.)

3.17 **Chemical enhancement**: The use of chemicals that react with specific types of evidence (e.g., blood, semen, lead, fingerprints) in order to aid in the detection and/or documentation of evidence that may be difficult to see.

3.18 **Chemical threat**: Compounds that may pose bodily harm if touched, ingested, inhaled, or ignited. These compounds may be encountered at a clandestine laboratory, or through a homemade bomb or tankard leakage (e.g., ether, alcohol, nitroglycerin, ammonium sulfate, red phosphorus, cleaning supplies, gasoline, or unlabeled chemicals).

3.19 **Circumstantial (Indirect) Evidence**: Evidence which indirectly proves a fact in issue. Requires the fact finder to use inferences (a conclusion that can be drawn from a fact.)

3.20 **Clean/sanitize**: The process of removing biological and/or chemical contaminants from tools and/or equipment (e.g., using a mixture of 10-percent household bleach and water).

3.21 **Collect/collection**: The process of detecting, documenting, or retaining physical evidence.

3.22 **Comparison samples**: A generic term used to describe physical material/evidence associated with crime scenes that may be compared with samples from persons, tools, and physical locations. Comparison samples may be from either an unknown/questioned or a known source.

**Samples whose source is unknown/questioned are of three basic types:**

1. **Recovered crime scene samples** whose source is in question (e.g., evidence left by suspects, victims).

2. **Questioned evidence** that may have been transferred to an offender/victim/other individual/ or item during the commission of the crime and taken away by him or her. Such questioned evidence can be compared with evidence of a known source and can thereby be associated/linked to a person/vehicle/tool of a crime.

3. **Evidence of an unknown/questioned source** recovered from several crime scenes may also be used to associate multiple offenses that were committed by the same person and/or with the same tool or weapon.
Samples whose source is known are of three basic types:

1. **A standard/reference sample** is material of a verifiable/documented source which, when compared with evidence of an unknown source, shows an association or linkage between an offender, crime scene, and/or victim/individual/location/or item (e.g., a carpet cutting taken from a location suspected as the point of transfer for comparison with the fibers recovered from the suspect's shoes, a sample of paint removed from a suspect vehicle to be compared with paint found on a victim's vehicle following an accident, or a sample of the suspect's and/or victim's blood submitted for comparison with a bloodstained shirt recovered as evidence).

2. **A control/blank sample** is material of a known source that presumably was uncontaminated during the commission of the crime (e.g., a sample to be used in laboratory testing to ensure that the surface on which the sample is deposited does not interfere with testing. For example, when a bloodstain is collected from a carpet, a segment of unstained carpet may be collected for use as a blank or elimination sample).

3. **An elimination sample** is one of known source taken from a person who had lawful access to the scene (e.g., fingerprints from occupants, tire tread impressions from police vehicles, footwear impressions from emergency medical personnel) and is generally used for comparison with evidence of the same type.

3.23 **Contamination**: The unwanted transfer of material from another source to a piece of physical evidence.

3.24 **Control/blank sample**: See comparison samples.

3.25 **Crime Scene Reconstruction**: The process of determining the nature and/or sequence of events that occurred at a scene from an evaluation of physical evidence and other relevant information observed at the scene.

3.26 **Cross-contamination**: The unwanted transfer of material between two or more sources of physical evidence.

3.27 **Datum Point**: A point which serves as a reference or base for the measurement of other quantities.

3.28 **Documentation**: Written notes, audio/videotapes, printed forms, sketches and/or photographs that form a detailed record of the scene, evidence recovered, and actions taken during the search of the crime scene.

3.29 **Dying declaration**: Statements made by a person who believes he or she is about to die, concerning the cause or circumstance surrounding his or her impending death.

3.30 **Elimination sample**: See comparison samples.
3.31 **Evidence identifiers**: Tape, labels, containers, and string tags used to identify the evidence, the person collecting the evidence, the date the evidence was gathered, basic criminal offense information, and a brief description of the pertinent evidence.

3.32 **First responder(s)**: The initial responding law enforcement officer(s) and/or other public safety official(s) or service provider(s) arriving at the scene prior to the arrival of the investigator(s) in charge.

3.33 **Fixative**: A spray or powder applied to an impression prior to chemical enhancement or casting.

3.34 **Impression evidence**: Objects or materials that have retained the characteristics of other objects that have been physically pressed against them.

3.35 **Initial responding officer(s)**: The first law enforcement officer(s) to arrive at the scene.

3.36 **Investigator(s) in charge**: The official(s) responsible for the crime scene investigation.

3.37 **Known**: See comparison samples.

3.38 **Latent print**: A print impression not readily visible, made by contact of the hands or feet with a surface resulting in the transfer of materials from the skin to that surface.

3.39 **Metal Detector**: A device used to detect metallic objects within a solid matrix.

3.40 **Multiple scenes**: Two or more physical locations of evidence associated with a crime (e.g., in a crime of personal violence, evidence may be found at the location of the assault and also on the person and clothing of the victim/assailant, the victim's/assailant’s vehicle, and locations the victim/assailant frequents and resides).

3.41 **Nonporous container**: Packaging through which liquids or vapors cannot pass (e.g., glass jars or metal cans).

3.42 **Oblique light**: Light that is positioned at a low angle of incidence relative to the surface being photographed or visualized. It is also referred to as side lighting.

3.43 **Other responders**: Individuals who are involved in an aspect of the crime scene, such as perimeter security, traffic control, media management, scene processing, and technical support, as well as prosecutors, medical personnel, medical examiners, coroners, forensic examiners, evidence technicians, and fire and rescue personnel.

3.44 **Personal protective equipment (PPE)**: Articles such as disposable gloves, masks, and eye protection that are utilized to provide a barrier to keep biological or chemical hazards from contacting the skin, eyes, and mucous membranes and to avoid contamination of the crime scene.

3.45 **Porous container**: Packaging through which liquids or vapors may pass (e.g., paper bags, cloth bags).
3.46 **Presumptive test**: A non-confirmatory test used to screen for the presence of a substance.

3.47 **Probable Cause**: Reasonable grounds or a substantial objective basis for believing that more likely than not an offense has been committed and a person to be arrested has committed it.

3.48 **Projectile trajectory analysis**: The method for determining the path of a high-speed object through space (e.g., a bullet emanating from a firearm).

3.49 **Radiological threat**: The pending exposure to radiation energy. (This energy can be produced by shortwave x-rays or through unstable isotopes.)

3.50 **Sampling selection** – A practice of selecting items to test, or portions of items to test, based on training, experience, and competence. In sample selection, there is no assumption about homogeneity.

3.51 **Scale**: An object showing standard units of length (e.g., ruler) used in photographic documentation of an item of evidence.

3.52 **Single-use equipment**: Items that are intended to be used only once to collect evidence, such as biological samples, then discarded to minimize contamination (e.g., tweezers, scalpel blades, droppers).

3.53 **Standard/reference sample**: See comparison samples.

3.54 **Taphonomy**: The study of processes which modify biological tissue within a forensic context. These include the contextual conditions in which remains are discovered, decomposition rates, environmental patterns, disarticulation and dispersion of human tissues, and the postmortem interval (time since death).

3.55 **Team members**: Individuals who are called to the scene to assist in investigation or processing of the scene (e.g., scientific personnel from the crime laboratory or medical examiner’s office, other forensic specialists, photographers, mass disaster specialists, experts in the identification of human remains, arson and explosives investigators, clandestine drug laboratory investigators, as well as other experts).

3.56 **Trace evidence**: Physical evidence that results from the transfer of small quantities of materials (e.g., hair, textile fibers, paint chips, glass fragments, gunshot residue particles).

3.57 **Track**: A mark left by footwear or a portion of a tire when it comes in contact with a receiving surface. The track may be two or three-dimensional depending on the nature of the substrate.

3.58 **Transient evidence**: Evidence which by its very nature or the conditions at the scene will lose its evidentiary value if not preserved and protected (e.g., blood in the rain).
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3.59 **Unknown/questioned:** See comparison samples.

3.60 **Walk-through:** An initial assessment that is conducted by carefully walking through the scene to evaluate the situation, recognize potential evidence, and determine resources required. Also, a final survey conducted to ensure the scene has been effectively and completely processed.

### 4.0 General Requirements

See Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual

### 5.0 Structural Requirements

See Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual
6.0 Resource Requirements

6.1. General

See Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual

6.2. Personnel

- If multiple forensic scientists respond to a scene, a lead shall be designated.
- Completion and accuracy of documentation and reporting shall be the responsibility of the lead.
- Non-lab personnel shall be kept to a minimum to ensure the integrity of the scene.

See Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual

6.3. Facilities and Environmental Conditions

Standard laboratory safety protocols should be followed in the field. Refer to the ACSO Forensic Lab Health and Safety Manual. See 6.3 in the ACSO’s Quality Assurance Manual, specifically section, 6.3.5, on activities performed outside of the Laboratory’s permanent control.

Ensure that the requesting deputy/officer understands they are responsible for providing scene security. Forensic scientists should not begin scene processing until security has been provided and should not continue processing the scene if reasonable security is not maintained.

To prevent contamination of personnel and the scene, the appropriate protective equipment should be utilized (e.g., gloves, Tyvek suits & shoe covers, dust masks, respirators, eye protection, etc.). To minimize cross-contamination, personal protective equipment should be worn and changed when necessary.

There shall be no smoking, eating, or drinking by personnel in the immediate vicinity of crime scenes or post-mortem examinations, except in designated safe areas.

Evidence collected for transport back to the laboratory should be packaged and sealed to maintain its integrity and prevent contamination of personnel or other items. Double paper bags or plastic bags may be used temporarily to contain items while they are in transit to the laboratory.

All evidence containing suspected biological fluids should be marked with appropriate biohazard labels. Firearms not already in a properly labeled firearms box should have firearm labeling/stickers to identify its contents.

Examination utensils (e.g., forceps, scissors, placards, measuring devices, etc.) used in crime scene processing should be placed in an appropriate container for subsequent disinfecting upon return to the laboratory.
6.4. **Equipment**

Equipment utilized for locating potential items/areas to test (e.g., flashlight, ALS, hand-held lens, and stereomicroscope) or documentation (e.g., camera and accessories) does not require routine calibration or maintenance.

6.4.1. **Alternate Light Source (ALS)**

The ALS can be used to detect a wide variety of forensic evidence using the principles of fluorescence, reflection, and absorption.

Although not considered test equipment, performance of the alternate light sources shall be evaluated annually by an internal analyst. This will include an assessment of the visibility of some commonly encountered body fluid stains, to include semen.

6.4.1.1. **Method**

Various items of forensic interest (e.g., trace evidence, biological stains) can be enhanced with the use of ALS with appropriate filters. Refer to the operations manual of the ALS model prior to use. Below are suggested wavelength and filter combinations for searching. Searching with other combinations may be warranted.

6.4.1.1.1. High intensity white light with clear/no filter and no goggles may be used to visualize blood. 415nm with orange goggles can assist looking for blood on patterned objects. It should be noted that blood absorbs the light and appears black or dark in color.

6.4.1.1.2. Violet (415-430nm) and blue (430-470nm) light colors with yellow/orange goggles can assist in visualizing biological substances.

6.4.1.1.3. The blue, blue-green (460-510nm) and green (500-550nm) light colors with orange and red goggles can assist in visualizing fibers and general trace evidence or fluorescent latent print powders and dyes.

6.4.1.2. **Cautions/Safety**

- Precautions should be used when operating any ALS.
- Proper eye protection shall be worn by anyone operating/within the area of the intense light source.
- Permanent eye damage can occur from direct illumination to the eye or reflected or refractive light hitting the eye.
- Exposing the skin to the beam of light can cause burns and otherskin damage.
6.4.2. **Metal Detector**

In some instances, a search for metal items may be needed. Metal items can include spent rounds, cartridge cases, and/or weapons. On surfaces such as concrete, asphalt, gravel, and hard dirt, the areas to be searched will most frequently be searched visually. In addition, the surface being searched may contain evidence at the sub-surface level. The use of a metal detector can be employed when searching any surface type, but would be most beneficial in the grassy, wooded, and overgrown areas or in instances when evidence may be located at a shallow sub-surface level.

6.4.2.1. **Methods**

Refer to manufacturer’s guide for general operation and maintenance information.

6.4.2.1.1. Verification of the metal detector shall be performed before each use.

Scatter a sampling of the following metals apart from each other and sweep the detector over each sample piece. The depth of the samples can be varied at the forensic scientist’s discretion.

a. Brass cartridge case  
b. Nickel-plated brass cartridge case  
c. Aluminum cartridge case  
d. Steel cartridge case  
e. Lead bullet  
g. Penny  
h. Iron  
j. Ground only (negative control – no sample)

6.4.2.1.2. Once the sensitivity is set; the forensic scientist is ready to use the metal detector for the search. The metal detector operates using an audible “beep” to alert the user of the possible presence of metallic objects within the area under the loop or coil. Some detectors emit a just audible background/threshold tone to aid in the location of targets.

6.4.2.1.3. As the forensic scientist walks the area of the search, the metal detector should be held to the front of the forensic scientist, and the loop or coil of the detector should be moved from left to right, back and forth, keeping the loop in continuous motion parallel to the ground being searched. If the loop or coil is held stationary, metal may not respond.

6.4.2.1.4. When an area being searched causes an audible “beep”, the loop of the detector should be trained on the area, moving the loop in an “X” pattern until the target is identified by finding the center of the “X” where the detector “beeps”.
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6.4.2.1.5. When the target area is determined, the forensic scientist should perform a visual search of the target area. The area should be thoroughly hand searched and, depending on the terrain, a sub-level search can also be performed.

6.4.2.1.6. After the area is hand searched, the forensic scientist should collect any items of evidentiary value using appropriate procedures for the handling and packaging of evidence. If no items of evidentiary value are located in the target area, the forensic scientist should continue to search the area, repeating the above procedure when audible “beeps” sound. Completion of the metal detector search is based on the parameters of the investigation and on the discretion of the forensic scientist.

6.5. **Metrological traceability**

There are no measurements taken by crime scene responders that require traceability.

All measurements are approximate, therefore measurement equipment (e.g. rulers, scales, etc.) does not require routine calibration or maintenance and it doesn’t need to be documented and/or tracked. Precise measurements are not necessary for reagent preparation as those utilized for testing are quality checked prior to use.

6.6. **Externally provided products and Services**

See Ada County Sheriff’s Office Forensic Crime Lab Quality Assurance Manual
7.0 Process Requirements

7.1. Review of request, tenders, and contracts

A request for service must be documented within the case documentation. Any deviation from the original request must also be documented. To see more information on review of request see the Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual.

7.2. Selection, verification, and validation of methods

See Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual

7.3. Sampling

Sample selection makes no inference about the entire population. Selection of items and samples is left to the analyst’s discretion and should be based on training and experience, while also considering the case circumstances. Sampling selection does not occur at crime scenes.

The decision of what evidence should be collected should be a joint effort between the forensic scientist and the detective(s). The proper collection and handling of evidence is the responsibility of the forensic scientist.

7.4. Handling of test or calibration items

If exigent circumstances exist during the investigation of a scene and it becomes obvious that items of evidence maybe perishable, lost, or destroyed (e.g., trace, biological fluid, and impression evidence), the forensic scientist should advise the lead investigator to determine if the evidence can and should be collected without consent to search or a search warrant, in order to preserve the perishable evidence.

See the Ada County Sheriff’s Office Forensic Lab Quality Assurance Manual for general evidence guidance. Items requiring specialized evidence packaging will be defined within discipline sections within the Crime Scene Analytical Method.

7.5. Technical Records

Crime scene documentation is extremely important to provide a detailed record of events, aid in report writing, assist with testimony, and allow for independent review by other experts.
7.5.1. **General**

7.5.1.1. Crime scene documentation includes a combination of notetaking, photography, and diagramming.

7.5.1.2. Documentation shall take place as crime scene processing is occurring, or shortly thereafter.

7.5.1.3. All original notes may be recopied. All original notes must be maintained as a permanent component of case records unless a legible and accurate copy is captured electronically.

7.5.1.4. If multiple lab staff respond to a scene, each forensic scientist shall document the tests they performed, reagent lot numbers used, controls, specific areas or items tested, and the equipment they utilized (e.g. Phenolphthalein testing, metal detector). This can be done by each forensic scientist taking notes or by one forensic scientist taking notes and the other forensic scientist(s) initialing those notes. Only the forensic scientist designated as lead for the scene needs to produce a report.

7.5.2. **Initial Response Request Documentation**

7.5.2.1. Document date of the request for scene processing and who made the request.

7.5.2.2. Document specific information provided regarding the scene and the source of the information.

7.5.2.3. Ascertain whether legal authority for processing has been obtained or is needed.

7.5.2.4. Attempt to determine if any specialized services or equipment may be required.

7.5.2.5. Obtain contact information for an investigator at the scene, if not known.

7.5.2.6. Document all requested work to be performed. Document any deviations from that request and communication regarding the deviation.

7.5.3. **Preliminary Scene Information**

7.5.3.1. Document the time of arrival at the scene and who is present.

7.5.3.2. Document that the scene has been secured appropriately.

7.5.3.3. The following information shall be included, if available:
   - Case number
   - Requestor
   - Scene location
   - Victim and suspect information
   - Other personnel present at the scene

7.5.3.4. For investigations conducted under a search warrant, review the warrant and ascertain the legal limits of the search. If available, retain a copy of the warrant.
7.5.4. Note-taking

Crime scene notes provide a thorough and comprehensive written account of actions, procedures, and analyses at the scene. Notes can include photographs and/or diagrams. Extensive notetaking shall be maintained throughout the scene investigation and may include:

- Scene condition: doors and windows locked/unlocked, opened/closed, lights on/off, shades up/down, temperature/weather conditions, odors, etc.
- General condition of victim(s)/individual(s): position of the body, apparent wound(s) present, clothing, personal belongings.
- Description and location of evidence (e.g., weapons, bloodstain patterns, fingerprints).
- Test(s) performed, and equipment utilized.
- Time of body removal and by who.

7.5.4.1. Diagramming

Crime scene diagrams serve to establish spatial relationships, provide an overall scene view, assist with preparation of demonstrative aids for court and serve as an investigative aide during interviews. In addition, diagrams can clarify items of evidence in a crime scene without extraneous items such as furniture, piles of debris, etc.

Types of diagrams for notes:

- Bird’s Eye View (Overview) Diagram:
  A sketch on one horizontal plane that shows the scene as if viewed from above.
- Exploded Diagram:
  Overview diagram with the addition of walls folded outward.
- Three-Dimensional (Perspective) Diagram
  A sketch depicting objects of evidence as they would appear to the eye with reference to relative distance, depth, and breadth.
- Elevation (Vertical Plane) Diagram
  A sketch on one vertical plane that shows the scene as viewed from the side.
- Forensic mapping devices, examples are Global Positioning Systems (GPS) and laser and electronic measuring devices (e.g., Total Data Station, Leica Disto).
### 7.5.4.2. Methods of Measurements for Documentation

When the forensic scientist is recording investigative data regarding distances/heights, measurements should be taken to provide a reference of the dimensions, to show the relationships of objects, and when necessary, properly document to enable scale reproductions.

**Types of Measurements:**

- **Rectangular Coordinates (Coordinate Method):**
  Two measurements at right angles are made from fixed objects, such as walls, to the item. Indicate the location of the fixed object and direction measured. Examples: “5 ft. east of south wall”, “4 ft. 6 in. south of north wall”, etc.

- **Triangulation:**
  Measurements are taken between two fixed objects and then from the fixed objects to the item, forming a triangle.

- **Polar Coordinates:**
  Measure from a fixed object (i.e. the building) to the item. Then measure the angle in a clockwise direction between the measuring line and a line through the fixed object.

- **Base Line Measurements:**
  Lay a tape down so that it crosses the entire room or area to be measured. Establish a reference point at each end of the tape, designated by a number or letter. The tape which runs between the two reference points becomes the base line for all other measurements in that area. Measurements are then made from the tape (baseline) by laying another tape measure perpendicular to the baseline out to the item of evidence or point of interest.
7.5.4.3. **Photography and Imaging**

7.5.4.3.1. **Photography Equipment**

Digital SLR camera (does not require calibration/maintenance), video camera, batteries (alkaline, lithium), memory cards or other digital storage media, external flash units for digital cameras, camera accessories to include cords, filters, shutter releases, and lenses, bubble level, scale/measuring device, and tripod.

7.5.4.3.2. **General Scene Photography**

To properly document a crime scene, overall, mid-range, and close-up photographs should be taken.

- **Overall Photographs:**
  Overall evidence should be used to establish where an item is located within the scene. To show the relationship of all evidence in an area or document the location of a crime scene. An example would be a photograph of the outside of a house with the street address showing or the living room inside the house where the body and a variety of different evidence is located. Some areas may require overlapping photos from each corner of the room. If placards are utilized to identify the location of evidence at the scene, photos should be taken before and after their placement.

- **Mid-range Photographs:**
  These photographs should begin to document the relationship of smaller evidence to each other within the overall scene.

- **Close-up Photographs**
  These pictures detail the individual item of evidence. They will document the condition or placement of the object or show evidence related to the crime. Close up photographs should be done with and without a scale. Close up photographs should also fill the frame. These photos should be used for identification purposes.

During scene processing, the entire scene should be photographed prior to collection of evidence or disturbing the scene. The following photographs should also be included, as appropriate.

- 360° perimeter view of the overall scene
- Scene location established by landmarks, street signs, addresses and adjacent areas
- All access routes to/from the scene
- All entrances/exits to the structure(s)
- Interior overall views with an overlapping series for each room (include ceilings, doorways, hallways, etc.)
- Intermediate and close-up views of individual items of evidence, continuity should be maintained between intermediate and close-up views.
Images of the scene used to depict a true and accurate representation of what was observed and processed (overall, mid-range and close-up images) should typically be acquired as either TIFF or the highest resolution JPEG format for the camera used. It is allowable to utilize another image capture system such as a mobile phone.

Upon return to the laboratory, images shall be downloaded to the secure imaging system(s) (LIMS/RIMS). Verify the number of images or the total size of data transferred to the imaging system(s).

7.5.4.3.3. **Crime Scene Video**
Videotaping may be utilized as an additional method of documenting the scene. When video documentation is performed at a crime scene, the forensic scientist should videotape the scene in the same manner as is utilized in still photography.

Landmarks will be used to identify the location of the scene. Evidence will be recorded in such a manner as to show spatial relationship of the item in context with other items or areas in the overall scene. Whenever possible, the sound should be muted during video recording.

Any video recorded will be turned over to a detective assigned to the case for retention or be booked into property/evidence.

7.5.4.3.4. **Examination Quality Photography**
Toolmarks, tire tracks, footwear impressions, bloodstain patterns, bite marks, friction ridge detail, injuries to live or deceased victims, and so forth are examples of evidence that may be found at a scene and in which accurate measurement and size documentation may be important to the investigation.

A scale shall appear in the photograph in order to produce an examination quality photograph print. Although the presence of a scale does not deem a photograph an examination quality photograph. Scales can and are used for relative sizes and distance determinations. If the purpose of the scale is for relative size only, a forensic examiner should document this in the case notes, if it’s not obvious in the photographs.

Examination quality images should also be taken at 90° (if possible) and in either the highest resolution jpeg or a lossless format.

Images shall be labeled in photographs or annotated in the digital imaging system.

7.5.4.3.5. **Luminol/BLUESTAR® FORENSIC Photography**
Viewing and photographing Luminol or Bluestar® reactions require the same environmental preparations to reduce or eliminate available light. Indoor scenes should be darkened as much as possible. Aluminum foil or other light blocking material may be necessary to cover windows and doors. Outdoor scenes should be photographed at night with as few
lights illuminated as possible. A tripod will need to be used to properly photograph the Luminol/Bluestar® low light conditions.

Utilize standard low light photographic techniques such as ISO, aperture setting, shutter speed. A suggested starting setting of f4.5 and 30 second exposure should be sufficient.

Note that the chemiluminescence reaction with BLUESTAR® can be bright enough to be photographed without significantly darkening the room; however, longer exposure times will typically be necessary to assure quality so use of a tripod is essential.

7.6. Testing Methods

Crime scene investigations are unique and require different approaches. Each scene requires a different combination of techniques, methods, flexibility, delegation of tasks, communication, and time management.

Positive and negative controls shall be documented for all test methods conducted at a scene with documentation of the results, when applicable.

7.6.1. Initial Scene Response, Assessment and Search

Crime scenes entail searching to ensure the recognition, documentation, collection, and preservation of all existing physical evidence. The recovered physical evidence may be used to include/exclude individuals, include/exclude suspects, and describe the objects and actions involved in the event. A forensic scientist should do this by doing the following steps:

• Conducting a systematic and thorough search of the scene. It is necessary for the forensic scientist to identify and collect items of evidence.
• Establish a path of entry to be used by all personnel who must enter the scene.
  Typically, this path should be a different route than that likely used by the perpetrator.
• Conduct a walk-through of the scene and document observations, with limited personnel.
  Personnel allowed within the scene at any time should have specific duties and should be kept to a minimum.
• Identify, protect, and document potential transitory evidence as soon as possible.

Apply appropriate search technique(s) to identify items of evidence. The following are examples of types of search techniques that may be applied.

• Spiral/Circle Method
• Strip Method
• Line Search
• Logical Association Method
After the initial search of the scene, discuss with the lead investigator the proposed method of processing. During discussions consider the following:

- Safety and health hazards
- The need for additional forensic scientists from specific disciplines
- The need for specialized resources (total station, anthropologist, aerial photographer, search teams, digital evidence specialist, etc.)
- Modification of scene boundaries, if necessary
- Personnel assignments (photography, scenediagram, evidence custodian, etc.)

Establish a staging area for equipment and supplies. This area should be located outside the actual scene or in an area that will preclude contamination.

7.6.2. Biological Evidence

Biological evidence encountered at a crime scene is generally in the form of liquid or dried blood, semen, and saliva. Other biological evidence that may be encountered are hair, urine, feces, bones, teeth and other tissues. Since biological evidence may lead to the identification of a victim or suspect, it is imperative that this evidence be collected and preserved for DNA analysis. Proper collection includes preventing contamination from extraneous sources such as crime scene personnel and other samples from the crime scene. Proper preservation ensures that degradation due to bacteria, humidity, high heat and other environmental factors is limited.

Photographs shall be taken prior to any testing, collection, or altering of the evidence.

7.6.2.1. Biological Evidence Collection

A description of the biological evidence should be documented (e.g., color, pattern, size, wet, clotted, damp or dry). Biological evidence should be packaged in paper (porous) products. The following order of preference for collection of biological evidence should be considered:

- Collect the entire item on which the evidence is located.
- Cut the evidence from its location. This is a reasonable course of action when dealing with large furniture, area rugs and carpet, drywall, door, and window frames, etc. Prior authorization (from Detectives, homeowners, etc. before destruction of property in some cases) should be obtained before any cuttings are collected on scene.
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- Swab the evidence. Sterile cotton swabs are the preferred method of collection. For dried stains, moisten a sterile swab using only enough water to collect the sample. For already damp or for wet stains the water step can be eliminated. Rotate the tip of the swab through the sample until adequate material has been collected. Concentrate the sample on the tip of the swab. Always consider collecting multiple swabs if sample quantity permits. Swabs from the same sample can be packaged together.

- In the case of liquid biological evidence, saturate sterile cotton swabs and allow to dry.

Special circumstances: Wet biological evidence should be dried prior to being placed into a storage location. It may be necessary to place wet biological evidence into plastic packaging for transportation purposes; however, it should be removed from the plastic as soon as possible, allowed to dry and then packaged in appropriate paper (porous) packaging. Visible trace evidence should be collected from these items before they are bagged and transported.

Presumptive blood testing may be performed at the scene, other biologic evidence should be submitted for consultation/testing with biology analyst after collection.

7.6.2.2. **Physical Examination**

Prior to chemical testing for biological evidence, physical examination shall be conducted. This can be done using one of the following techniques:

**High Intensity Light:** When searching a crime scene for blood, the forensic scientist shall first use a high intensity light. Diluted blood will often leave a brownish stain. Blood may flow into floorboard cracks, carpet padding and behind baseboards. Blood in these types of areas may be located with a high intensity light source and presumptive blood testing can be performed.

**ALS:** Many biological substances, such as semen, saliva, and urine will fluoresce under certain wavelengths of light and goggle/filter (red, orange, yellow, clear/UV protection) combinations. Visualization of biological fluids is typically optimal near 450 nm; however, it is recommended to test other wavelengths to determine the optimal contrast. The ALS is a search tool and does not indicate the identification of any substance. Additional testing is required when fluorescence is detected. Fluorescence along with crime scene knowledge is used to determine further testing procedures at the crime scene. Any detection/search shall be documented in the case notes.
7.6.2.3. **Presumptive Blood Testing/Enhancement**

7.6.2.3.1. **BLUESTAR® FORENSIC**

Bluestar® is a catalytic blood test that gives a positive reaction in the presence of blood due to the peroxidase activity of hemoglobin. A positive reaction is a chemiluminescence that is observable in a dark environment. Bluestar® is most useful when blood is suspected but is not visible and may be used to help locate non-visible blood stains. Bluestar® is an ideal reagent to locate non-visible blood or enhance suspected blood that is on black or dark colored porous and non-porous substrates. More specific presumptive blood tests, such as phenolphthalein, may be subsequently used on Bluestar® positive areas/stains.

**Supplies**

**BLUESTAR®**

For the BLUESTAR® FORENSIC tablets you will need distilled water & a 125 ml spray bottle (mister) equipped with an adjustable spray nozzle.

**Method**

The working solution is prepared before use. For optimal results, this solution should be used within 3-4 hours after mixing. Each BLUESTAR® Forensic tablet reagent contains a beige tablet (containing Sodium Hydroxide) and a white tablet (containing Hydrogen Peroxide – Urea) which will make 125 ml of solution. 125 ml is generally sufficient for a 250 sq. ft. area. Photography techniques should be prepared prior to the application/preparation of BLUESTAR® FORENSIC to ensure the forensic scientist is ready to capture any reaction.

- Remove the two tablets from the reagent packet and add to 125 ml of distilled water.
- Allow to completely dissolve (~1 to 2 minutes).
- Gently stir with a circular motion. DO NOT shake the container upside down.
- Determine the area to be sprayed with BLUESTAR®.
- Lightly mist the area and photograph immediately after application.
- Additional application may be necessary. However, use caution to avoid saturation and dilution of the target.
- Samples should be collected from the area(s) which displayed luminescence for additional testing.

**Photography**

Utilize standard low light photographic techniques such as ISO, aperture setting, shutter speed, and subject lighting. A suggested starting setting of f4.5 and 30 second exposure should be sufficient.

Note that the chemiluminescence reaction with BLUESTAR® can be bright enough to be photographed without significantly darkening the room;
however, longer exposure times will typically be necessary to assure quality so use of a tripod is essential.

Viewing and photographing Luminol or BLUESTAR® reactions require the same environmental preparations to reduce or eliminate available light. Indoor scenes should be darkened as much as possible. Aluminum foil or other light blocking material may be necessary to cover windows and doors. Outdoor scenes should be photographed at night with as few lights illuminated as possible. A tripod will need to be used in order to properly photograph the Luminol/BLUESTAR® low light conditions.

**Interpretation**

**Positive:** Chemiluminescence with a color and persistence like the positive control indicates the possible presence of blood.

**Negative:** No chemiluminescence, chemiluminescence that differs from the positive control in appearance or duration, or chemiluminescence lacking an apparent pattern or discernable area of distinction (i.e., entire item/area luminesces) does not indicate the presence of blood.

**Inconclusive:** An atypical reaction or other circumstance that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to this determination.

**Cautions/Safety**

- All other analysis and search options should be considered prior to BLUESTAR® application.
- Spraying of BLUESTAR® dilutes bloodstains.
- False Positive: False chemiluminescence reactions may occur with the presence of certain household detergents, chlorine, some paints and varnishes, copper, and certain iron metabolizing plants and soils containing iron.
- Such “false” reactions may be identifiable because their color, brightness, and duration differ from those of the typical reaction with blood. False positives may result in a whitish chemiluminescence or one that has a fast-initial burst of color that rapidly diminishes.
- Because the BLUESTAR® reagent is primarily water it will cause dried bloodstains to dissolve. Any pattern evidence (e.g., shoeprints, fingerprints, impact spatter, etc.) will be negatively affected. If pattern evidence is located, BLUESTAR® use should be immediately stopped and the evidence and scene assessed to determine the best method to use in proceeding.
- BLUESTAR® works with blood stains and impressions. Consider testing with a more specific presumptive test for blood prior to application, when possible.
- Any samples collected after BLUESTAR® processing should be thoroughly dried prior to packaging. If possible, sample prior to the use ofBLUESTAR®.
- Photographic equipment should be set up and ready for use prior to
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BLUESTAR® application.
• BLUESTAR® is a relatively non-specific presumptive test for blood, so other substances such as chlorine, metallic ions like manganese, copper and iron, some paints and varnishes, some vegetables, fruits, etc. may yield a positive reaction.
• Use in a ventilated area when possible.
• Wear eye protection and masks when spraying any reagent.

Controls
Controls are checked prior to use.
Positive Control: A known blood sample on a substrate or swab/filter paper.
Negative Control: Unstained substrate or swab.

Documentation
The Lot number shall be documented in the case notes and expiration.

BLUESTAR® has a short shelf life and therefore need to be made whenever it is used. Therefore, the date of expiration is the date it is mixed up.

The results for both the casework, the positive control, and negative control shall be documented in the case notes.
7.6.2.3.2 **Luminol**

Luminol is used on suspected blood samples to locate non-visible blood or to enhance bloody impressions. Luminol is a clear, colorless reagent. When luminol comes into contact with hemoglobin an oxidation reaction, catalyzed by the peroxidase-like activity of hemoglobin, occurs. A positive result is a temporary light blue chemiluminescence visible best in a dark environment. The resulting chemiluminescence may increase the contrast of a blood impression on a dark substrate.

Luminol is especially useful for suspected blood deposited on dark colored porous or non-porous surfaces. Results may be best on faint impressions or samples.

More specific presumptive blood tests, such as phenolphthalein, may be subsequently used on luminol positive areas/stains.

**Reagent**

The reagent is mixed just prior to use.

- Sodium carbonate, anhydrous ≥99.0% 10 gm
- Luminol > 97.0% 0.2 gm
- Deionized water 200 ml
- Sodium perborate monohydrate 1.4 gm

Dissolve the sodium carbonate, sodium perborate, and luminol in the 200mL deionized water. Complete dissolution may take several minutes at room temperature. Place the reagent in a non-metal fine-mist sprayer for application.

Luminol is considered expired within 1-2 hours of preparation or when the controls don’t yield the expected results, whichever comes first.

**Method**

- Determine area to be sprayed with Luminol. Application must occur in a dark environment to view any resulting chemiluminescence.
- Have a camera on a tripod or stabilized ready to document any positive reaction.
- Mix reagent and test controls. Document lot numbers and control reactions.
- Lightly spray the area with Luminol and photograph any positive reactions.
- Repeat spraying may be necessary. However, use caution since this will dilute the stain and may cause running.
- Samples may be collected as needed from the positive area(s) for additional testing and/or preservation.
Photography
Utilize standard low light photographic techniques such as ISO, aperture setting, shutter speed, and a tripod. A suggested starting setting of f4.5 and 30 second exposure should be sufficient.

Interpretations
Positive: Chemiluminescence with a color and persistence like the positive control indicates the possible presence of blood.

Negative: No chemiluminescence, chemiluminescence that differs from the positive control in appearance or duration, or chemiluminescence lacking an apparent pattern or discernable area of distinction (i.e., entire item/area luminesces) does not indicate the presence of blood.

Inconclusive: An atypical reaction or other circumstance that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to this determination.

Cautions/Safety
• Blood impressions may be dissolved by the luminol reagent causing running or obliteration of fine detail. This is more likely with repeated spraying.
• False Positive: False chemiluminescence reactions may occur with the presence of certain household detergents, chlorine, some paints and varnishes, copper, and certain iron metabolizing plants and soils containing iron.
• Such “false” reactions may be identifiable because their color, brightness, and duration differ from those of the typical reaction with blood. False positives may result in a whitish chemiluminescence or one that has a fast-initial burst of color that rapidly diminishes.
• Applying a chemical blood fixative (e.g. 2% 5-sulfosalicylic acid solution) prior to luminol treatment will reduce the luminol reaction and is not recommended.
• Any samples collected after luminol processing should be thoroughly dried prior to packaging. If possible, sample prior to the use of luminol.

• Photographic equipment should be set up and ready for use prior to luminol application.
• Upon drying, luminol contaminates the substrate with its component chemicals leaving a white crusty residue.
• Other analysis and search options should be exhausted prior to luminol application.
• Consider testing with a more specific presumptive test for blood prior to application when possible. Luminol is a relatively non-specific presumptive test for blood. Other substances such as chlorine, rust, iron-containing soil, metals, etc. may yield a positive result.
• Use in a ventilated area when possible.
• Wear eye protection and masks when spraying any reagent.
Controls
Controls are checked prior to use to verify the reagent is working.
Positive Control: Known blood on a substrate or swab/filter paper.
Negative control: Unstained substrate

Documentation
The Lot number shall be documented in the case notes and expiration.
Luminol has a short shelf life and therefore need to be made whenever it is used. Therefore, the date of expiration is the date it is mixed up.
The results for both the casework and the positive and negative control shall be documented in the case notes.
7.6.2.3.3. **Phenolphthalein**

This oxidative test for the presumptive identification of blood is based on the catalytic activity of the heme group within the hemoglobin molecule. The phenolphthalein test is extremely sensitive and can be used to detect visible blood or blood that has been diluted or washed away. It may be commercially purchased.

**Reagents** (may be commercial purchase)

**Stock Solution**

<table>
<thead>
<tr>
<th>Phenolphthalein</th>
<th>2 g</th>
</tr>
</thead>
<tbody>
<tr>
<td>Potassium hydroxide</td>
<td>20 g</td>
</tr>
<tr>
<td>Deionized water</td>
<td>100 ml</td>
</tr>
<tr>
<td>Zinc, granulated, particle size approximately 20 mesh</td>
<td>20 g</td>
</tr>
</tbody>
</table>

Reflux the above solution until it becomes colorless (2-3 hours). Store in refrigerator in amber bottle with additional zinc added.

**Working solution**

<table>
<thead>
<tr>
<th>Stock solution</th>
<th>1 part</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ethanol</td>
<td>4 parts</td>
</tr>
</tbody>
</table>

Store in dropper bottle with mossy or granulated zinc.

(Commercially purchased Phenolphthalein requires added drops of ethanol in the testing method due to no added ethanol in the solution).

3% Hydrogen peroxide

**Method**

- Take a small cutting/swab or other transfer method of the targeted stain.
- If using a commercial reagent that does not contain ethanol, apply 1 to 2 drops of ethanol.
- Place 1 to 2 drops of phenolphthalein working solution or commercial phenolphthalein reagent on the cutting or swab. Allow the reagent to soak into the sample.
- Place 1 to 2 drops of 3% hydrogen peroxide on the sample. Interpret and document the results in the case notes.
Interpretation
Positive: A positive reaction will show a pink color within 10 seconds after the addition of working solution B (hydrogen peroxide) indicating the presence of blood.

Negative: No color change within 10 seconds indicates blood is not present or is too limited in quantity to be detected.

Inconclusive: A positive reaction that occurs prior to the addition of the hydrogen peroxide, development of a different color, or other circumstances that lead the forensic scientist to be cautious about a positive or negative interpretation. Note: the forensic scientist must document in their case notes the observation(s) that led them to make an inconclusive determination.

Cautions/Safety
• Do not place reagents directly on the evidence. Sample the evidence with a cutting or swabbing.
• It should be noted that the activity of the 3% hydrogen peroxide solution decreases over time. Caution should be used when using hydrogen peroxide that is older than 6 months.
• To optimize the efficacy of hydrogen peroxide, forensic scientists must store the working solution in a refrigerator/cooler when not in use and limit exposure to air and light.
• In the absence of blood, the two reagents will begin to react with each other and give a pink color with time.
• The phenolphthalein test is a presumptive test and substances other than blood may yield positive reactions.

Controls
Controls are checked prior to use. A known documented sample from the lab is used to verify the control is working.

Positive: A known blood stain.

Negative: Unstained swab, fabric, filter paper, or empty well.

Documentation
The Lot number(s) shall be documented in the case notes.

The results for both the casework and the positive and negative control shall be documented in the case notes.
7.6.2.3.4. **Leucomalachite Green (LMG)**

This oxidative test for the presumptive identification of blood is based on the catalytic activity of the heme group within the hemoglobin molecule. The leucomalachite green test is extremely sensitive and can be used to detect visible blood or blood that has been diluted or washed away.

**Reagents** (may be a commercial purchase)

<table>
<thead>
<tr>
<th>Substance</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leucomalachite green</td>
<td>0.06 g</td>
</tr>
<tr>
<td>Sodium perborate</td>
<td>0.2 g</td>
</tr>
<tr>
<td>Methanol</td>
<td>20 mL</td>
</tr>
<tr>
<td>Glacial acetic acid</td>
<td>10 mL</td>
</tr>
</tbody>
</table>

Reagent should be prepared with each use.

**Method**

- Take a small cutting/swab or other transfer method of the targeted stain.
- Place 1 to 2 drops of leucomalachite green mixed reagent or commercially purchased reagent on the cutting or swab. Allow the reagent to soak into the sample.
- Interpret and document the results in the case notes.

**Interpretation**

Positive: A positive reaction will show a dark green color almost immediately (10 seconds).

Negative: No color change within 10 seconds.

Inconclusive: The development of a different color or other circumstances that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination.

**Cautions/Safety**

The leucomalachite green test is a presumptive test and substances other than blood may yield positive reactions.

**Documentation**

The Lot number and shall be documented in the case notes.

The results for both the casework and the positive and negative control shall be documented in the case notes.

**Controls**

- Controls are checked prior to use.
- Positive: A known blood stain.
- Negative: Test an unstained swab, fabric, filter paper, or empty well.
7.6.2.3.5. **O-Tolidine (O-tol)**

This oxidative test for the presumptive identification of blood is based on the catalytic activity of the heme group within the hemoglobin molecule. The o-tolidine test is extremely sensitive and can be used to detect visible blood or blood that has been diluted or washed away.

**Reagents**

<table>
<thead>
<tr>
<th>Reagent</th>
<th>Quantity</th>
</tr>
</thead>
<tbody>
<tr>
<td>O-tolidine</td>
<td>0.6 g</td>
</tr>
<tr>
<td>Glacial acetic acid</td>
<td>100 mL</td>
</tr>
<tr>
<td>Ethanol</td>
<td>100 mL</td>
</tr>
<tr>
<td>3% Hydrogen Peroxide</td>
<td>1-2 drops</td>
</tr>
</tbody>
</table>

Dissolve o-tolidine in acetic acid/ethanol mixture.

**Method**

- Take a small cutting/swab or other transfer method of the targeted stain.
- Place 1 to 2 drops of o-tolidine reagent on the cutting or swab. Allow the reagent to soak into the sample.
- Place 1 to 2 drops of hydrogen peroxide on the sample.
- Interpret and document the results in the case notes.

**Interpretation**

Positive: A positive reaction will show a blue/green color within 10 seconds after the addition of hydrogen peroxide.

Negative: No color change within 10 seconds.

Inconclusive: A positive reaction that occurs prior to the addition of hydrogen peroxide, development of different color, or other circumstance that lead the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to the inconclusive determination.

**Cautions/Safety**

- Do not place reagents directly on the evidence. Sample the evidence with a cutting or swabbing.
- It should be noted that the activity of the hydrogen peroxide solution decreases over time. Caution should be used when using hydrogen peroxide that is older than 6 months.
- In order to optimize the efficacy of hydrogen peroxide, forensic scientists must store the reagent in a refrigerator/cooler when not in use and limit exposure to air and light.
- The o-tolidine test is a presumptive test and substances other than blood may yield positive reactions.
- In the absence of blood, the two reagents will react with each other and eventually develop a blue-green color over time due to the
nature of oxidation reactions. Thus, it is important to adhere to the allotted reaction times.

Controls
Controls are checked prior to use. Working solutions are checked at the time of use.

Positive: A known blood stain.
Negative: Unstained swab, fabric, filter paper, or empty well.

Documentation
The Lot number shall be documented in the case notes.

The results for both the casework and the positive and negative control shall be documented in the case notes
7.6.2.3.6. **Amido Black – Methanol Based**

Amido black is used on bloody impressions and is a dark blue-black colored reagent. It reacts with the proteins present in blood, staining blood a blue-black color. The background reagent can be rinsed away which may increase the contrast of the impression on the substrate.

Amido black is especially useful for bloody or protein-based impressions deposited on lighter colored, non-porous surfaces such as glass, plastic, vinyl, etc. It may also work on concrete or some papers. Results may be best on faint impressions.

**Reagents**

**Developer Solution**

- Naphthol blue black: 2 g
- Glacial acetic acid: 100 mL
- Methanol: 900 mL

Dissolve the naphthol blue black in the above ingredients.

**Rinse Solution**

- Glacial acetic acid: 100 mL
- Methanol: 900 mL

Optional Rinse – Rinse with distilled water.

The Reagent and Rinse solutions are stored in bottles at room temperature or refrigeration.

The Reagent and Rinse solutions do not expire and may be used until the entire volume is consumed.

**Method**

- Use Reagent and Rinse solution on a small area of substrate that is not part of the impression to check for background staining. Do not use this reagent if significant background staining occurs.
- Apply the Reagent to the impression via spraying, pouring, or submersion.
- To ensure complete staining, the solution should remain in contact with the impression for at least 1-2 minutes to obtain maximum development.
- Apply the Rinse solution to remove Reagent stain from background areas. An optional water rinse may follow.
- Allow the impression to air dry.
- The impression may be re-stained to make darker, if desired.

**Interpretation**

Positive: A blue-black staining will appear within 1-2 minutes.
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Negative: No change or less intense blue-black staining results.

Inconclusive: Any area that development occurs of a different color or other circumstances that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination

Cautions/Safety
• Use in a very well-ventilated area. Methanol and glacial acetic acid in the quantity and concentration of this formulation are inhalation hazards.
• Wear eye protection and masks when spraying any reagent.
• Amido black works only with bloody or other protein-based impressions. Consider a presumptive test for blood prior to application, when possible.
• Porous substrates may absorb the blue-black reagent causing background staining that cannot be rinsed away. This might result in little contrast with the treated bloody impression.
• Fresh blood impressions may be damaged or destroyed if not fixed prior to treating.

Controls
Controls are checked with the reagent prior to use.
Positive: A known blood stain on a substrate.
Negative: An unstained area of the substrate.

Documentation
The Lot number(s) shall be documented in the case notes.
The results for both the casework and the positive and negative control shall be documented in the case notes.
7.6.2.3.7. **Amido Black – Water Based**

Amido black is used on bloody impressions and is a dark blue-black colored reagent. It reacts with the proteins present in blood, staining blood a blue-black color. The background reagent is rinsed away which may increase the contrast of the impression on the substrate.

Amido black is especially useful for bloody or protein-based impressions deposited on lighter colored, non-porous surfaces such as glass, plastic, vinyl, etc. It may work on concrete or some papers. Results may be best on faint impressions.

The water-based formula may be considered for use when the substrate is sensitive to methanol or in any situation where a less vaporous solution is desired.

**Reagent**

**Developer Solution**

- Deionized water 500 mL
- 5-sulfosalicylic acid dihydrate, ≥99.0 20g
- Naphthol blue black 3g
- Sodium carbonate 3g
- Formic acid 50 mL
- Glacial acetic acid 50 mL
- Kodak Photo Flo solution 37.5 mL

Combine all reagent components in a ≥1L capacity bottle. Dilute mixture to 1L with deionized water. Although the mixture will be ready to use immediately, allow the mixture to stand for several days prior to use for best results.

The solution is stored in a bottle at room temperature or refrigeration. This reagent does not expire and may be used until the entire volume is consumed.

**Rinse** – water

**Method**

- Use developer solution and rinse solution on a small area of substrate that is not part of the impression, to check for background staining. Do not use this reagent if significant background staining occurs.
- Apply the Reagent to the impression via spraying, pouring, or submersion. To ensure complete staining, the solution should remain in contact with the impression for at least 3-5 minutes to obtain maximum development.
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- Rinse with water to remove Reagent stain from background areas. Allow the impression to air dry.
- The impression may be re-stained to make darker, if desired.

**Interpretation**

Positive: A blue-black staining will appear within 3-5 minutes

Negative: No change

Inconclusive: Any area that development occurs of a different color, or other circumstances that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination

**Cautions/Safety**

- Amido black works only with bloody or other protein-based impressions.
- Consider a presumptive test for blood prior to application, when possible.
- Porous substrates may absorb the blue-black reagent causing background staining that cannot be rinsed away. This might result in little contrast with the treated bloody impression.
- Use in a ventilated area when possible.
- Wear eye protection and masks when spraying any reagent.

**Controls**

Positive: A known blood stain on a substrate

Negative: An unstained area of the substrate

Controls are checked with the reagent prior to use.

**Documentation**

The Lot number(s) shall be documented in the case notes.

The results for both the casework and the positive and negative control shall be documented in the case notes.
7.6.2.3.8. **Leucocrystal violet (LCV)**

LCV is for use on bloody impressions. LCV is the reduced form of crystal violet and is a clear colorless reagent. When LCV and hydrogen peroxide come into contact with the hemoglobin in blood an oxidation reaction catalyzed by the peroxidase-like activity of the hemoglobin will occur. The result is a dark violet dye which has an affinity for proteinaceous substrates. The resulting violet color may increase the contrast of an impression on a substrate.

LCV is especially useful for bloody impressions deposited on lighter colored porous and non-porous surfaces such as vinyl flooring, carpeting, fabric, etc. Results may be best on faint impressions.

**Reagent**

- 5-sulfosalicylic acid dihydrate, ≥99.0: 10g
- 3% Hydrogen peroxide: 500 mL
- Sodium acetate anhydrous: 3.7g
- Leucocrystal violet: 1g

Combine the 5-sulfosalicylic acid, sodium acetate, and leucocrystal violet with the 500mL 3% hydrogen peroxide in a dark bottle. A 473mL volume of 3% hydrogen peroxide (the volume commonly sold at pharmacies) is an acceptable substitute for the 500mL 3% hydrogen peroxide volume.

The solution should be stored in a dark bottle and refrigerated. It expires 30 days after mixing.

**Optional rinse – water**

**Method**

- Apply LCV to the impression via spraying, pouring, or submersion and allow it to remain in contact with the impression for at least 30 seconds.
- Rinse non-porous substrates with water to remove excess LCV when necessary.
- Allow the impression to air dry.
- Enhanced impressions that aren’t rinsed should be photographed as soon as possible. This is to document the impression prior to any background color development that may occur.

An indirect sampling method may be used prior to LCV application, if desired. Define the area to be sampled and tested. Moisten filter paper with deionized water and press it onto the defined area. Mark the filter paper before removing it so the orientation is documented.
Remove the filter paper and spray with LCV then proceed with the method described above. This technique may be less sensitive than directly spraying, pouring, or submersion because it relies on the efficacy of the stain to transfer to the filter paper.

**Interpretation**

Positive: A violet color within 30 seconds to 3 minutes

Negative: No color change. Unreacted areas will also turn violet over time if not rinsed. See Cautions.

Inconclusive: Any area that development occurs of a different color, or other circumstances that causes the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination

Because impressions that are not visible may be enhanced or detected with the application of LCV, a violet color reaction may be interpreted and reported as a positive result with a presumptive test for blood.

**Cautions/Safety**

- LCV crystals that have turned yellow should not be used.
- The LCV reagent is light sensitive and may discolor over time due to exposure. LCV reagent should be colorless or near colorless. Should it appear blue or violet, consider making up fresh reagent. Store in dark bottles.
- Treated areas will change from colorless to violet over time unless rinsed. The timing varies with environmental conditions and may occur within hours to several days after application.
- Use in a ventilated area when possible. Wear appropriate eye protection when using.

**Controls**

Positive: A known blood stain on a substrate

Negative: An unstained area of the substrate

Controls are checked with the LCV and rinse prior to use.

**Documentation**

The Lot number and expiration date shall be documented in the case notes.

The results for both the casework and the positive and negative control shall be documented in the case notes.
7.6.2.3.9. 2% 5-sulfosalicylic acid solution

A 2% 5-sulfosalicylic acid solution fixes bloody impressions so they are not washed away during enhancement. Bloody impressions may fix through aging, but fresh blood impressions may be fixed with heat or a 2% weight/volume aqueous solution of 5-sulfosalicylic acid. A prior fixing step is not necessary if 5-sulfosalicylic acid is a component of the enhancement reagent.

**Reagent**

<table>
<thead>
<tr>
<th>Component</th>
<th>Quantity</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-sulfosalicylic acid dihydrate</td>
<td>20g</td>
</tr>
<tr>
<td>Deionized water</td>
<td>1 L</td>
</tr>
</tbody>
</table>

Dissolve the 20g 5-sulfosalicylic acid in 1L water.

The solution may be stored in bottles at room temperature or refrigerated. This reagent does not expire and may be used until the entire volume is consumed.

**Method**

The solution can be applied to the impression via spraying, pouring, or submersion. To ensure complete fixing, the solution should remain in contact with the impression for several minutes at which point excess solution is rinsed off with deionized water.

**Control**

This reagent cannot be reasonably subjected to a Quality Control test either by testing against a known standard prior to use or by end point verification; therefore, no control is necessary.
7.6.3. Impression Evidence

Impression evidence may take the form of a shoe, tire, toolmark, fabric, or any patterned impression. Wet or fragile impressions are susceptible to alteration during crime scene processing, so caution must be taken to preserve this type of evidence. Stepping Plates may be used to protect the floor surfaces during processing where impressions may be present.

7.6.3.1. Shoeprint Standard Evidence

Some scenes where shoeprint evidence has been collected will require the collection of elimination shoeprints from victims, suspects, witnesses, and other individuals associated with the scene. Collection of the pair of shoes is preferred, but not always reasonable especially with witnesses or emergency responders. There are several methods used to collect elimination shoeprint impressions.

- Dust the soles of the shoes with fingerprint powder and press the sole onto a sheet of adhesive paper. Place a clear sheet of plastic over the print to protect it.
- Lightly smear the outsoles of the shoes with petroleum jelly then press the outsoles onto a sheet of clean white paper. The impressions may be enhanced by lightly dusting with dark fingerprint powder.
- An inkless shoe print kit can be used by having the individual step onto the inkless pad and then onto the chemical sensitive paper.
- Other products available for use are gel lifters and Biofoam.

7.6.3.2. Chemical Enhancement Methods

The purpose of chemical enhancements is to increase the detail and contrast between an impression and its substrate. A forensic scientist, when authorized, may perform specific chemical enhancements on scene if necessary. Visible impressions or areas of suspected non-visible impressions that are not easily transported may be chemically enhanced on site. If impression evidence can be collected without damage, it shall be packaged and transported from the scene for chemical enhancement later in the laboratory under controlled conditions.

Impression evidence requiring chemical enhancement suspected of blood should utilize chemicals found in the presumptive blood test/enhancement. Specifically, forensic scientist(s) should utilize one of the following: LCV, Amido Black, Bluestar, and/or Luminol for impression enhancement if authorized on scene.

Examination quality images shall be taken of a visible impression before enhancement. Documentation of the impression’s location relative to its substrate and/or immediate surroundings shall also be included in the case notes.
After enhancement, examination quality images shall be taken of the impression if the enhancement resulted in greater detail, greater contrast, or visualization of additional impressions. If examination quality images are not taken after enhancement, the notes shall indicate the reason.

7.6.3.3. Lifting

Lifting involves transferring a two-dimensional impression from its original surface to a surface that will provide better contrast, allow easier transportation, and perhaps aid in preservation. Two methods are available for lifting impressions: electrostatic lifters and adhesive lifters. The examiner should consider the substrate (porous or nonporous, wet or dry), substrate condition (wet, dry), substrate color, and the presence of interfering material (dirt, grease) when selecting a lifting device. There is a risk that the impression may be destroyed during attempts to do the lift. Therefore, footwear impressions should only be lifted in the field when the item containing the impression cannot be safely transported to the laboratory or if a photograph is not sufficient.

7.6.3.4. Electrostatic dust lifting device

The electrostatic dust lifting device (ESDL) should be used on impressions of dry dust or dry residue on surfaces that are relatively clean. The device will work on both porous and non-porous substrates. If the impression is a wet residue or has become wet or damp prior to lifting, the ESDL may not work. The ESDL does not destroy wet prints. Therefore, if the type of impression is unknown the ESDL should be used first. If the print is not lifted, then subsequent method(s) can be used.

Equipment
- Electrostatic dust lifter
- Lifting film

Method
- Take examination quality photograph(s) of the impression prior to any lifting attempts.
- Assemble the ESDL. Attach the metal probe in the high voltage supply port if needed. Connect the ground wire, either antenna ground or alligator clip, to the ground port. Supply voltage to the main unit either with batteries or AC/DC outlet.
- Position the grounding device and attach the ground wire of the unit to the ground plate or other grounding material. The most effective position of the ground plate is when it is in maximum contact to a surface adjacent the impression.
- For impressions on moveable objects, such as newspapers, rugs, etc. the best position for the ground plate would be under the...
impression with the metal side up.

- For impressions on *immovable objects*, such as vinyl flooring or a wall, place the ground plate (metal side down) or the metal antenna next to, but not touching, the object.

- For impressions on *metallic objects*, such as a vehicle door, the ground wire may be attached directly to the metallic object.

- For metallic surfaces, first place a thin (1 or 2 mil), clear piece of Mylar or polyester on the impression before placing the lifting film. This step is not necessary for impressions on non-metallic surfaces.

- Place clean lifting film over the impression with the black side down (toward the impression) and silver side up (away from the impression). The lifting film must be larger than the impression to ensure a full transfer (it is prudent to use a larger piece than expected especially with latent or partial prints, as more detail may be present that is not apparent to the unaided eye).

- Mark the orientation of the film, if necessary.

- Place the probe on a corner of the metallized surface of the lifting film and turn on the voltage. Start at a lower voltage setting, and increase voltage as needed. It is not necessary to move the probe around the film. If any air bubbles develop as the film adheres to the substrate, they may be rolled out with a clean fingerprint roller.

- Discontinue the voltage to the film and allow the film to sit for at least 5 seconds to allow the static charge to dissipate.

- Remove the film, turn over on a clean flat surface, and evaluate the results. The results should be evaluated in a darkened room with strong oblique light.

- Subsequent lifts may be performed if the first was unsuccessful if due to a large quantity of transferred material (unsuccessful lifts due to an overall lack of transfer will not be successful in subsequent lift attempts). Second or third lifts may yield a “cleaner” impression as interfering background material may have been reduced during the first lift.

- A successful lift must be retained by securing it to prevent movement or destruction of the lift. Examination quality photographs are suggested. Unsuccessful lifts shall be noted but do not need to be photographed or maintained.

- The lifting film of unsuccessful lifts may be reused after cleaning with a soft cloth.

**Cautions/Safety**

- Not all dry impressions can be successfully lifted using ESDL.

- Attempts to lift dust prints on dirty backgrounds will cause both the dust print and dirty background to lift together. Subsequent lifts should be attempted to see if they would be successful.
• Never slide the lifting film across the surface with the impression.
• If using a fingerprint roller to eliminate air bubbles in the lifting film, do so gently to avoid shifting or excessive pressure that may damage the impression.
• As a precaution the electrostatic dust print lifter should not be operated by persons with pacemakers.
• A static electricity shock may occur when the forensic scientist touches the film if the forensic scientist fails to discharge the lifting film with the probe. To discharge the probe, allow it to sit in contact with the lifting film, with no voltage, for at least 5 seconds after the ESDL voltage is turned off.
• It is possible to receive electrical shocks from the lifting film, the ground plate, the metal probe, and a metallic surface. Such shocks will be avoided by not touching any of these parts when the current is on and by allowing the probe to discharge after use (see caution above).
• Do not allow the lifting film to come into contact with the grounding plate while the ESDL is on – this will cause arcing and the device will not work properly.
• If arcing occurs between the film and the ground, the power is too high, or part of the film is touching or too close to the ground plate.

**Documentation**

• Mark the lifting film before removing it from the surface with the impression to allow for orientation. This may be impossible to do if the film was removed from the surface without orientation marks, clear photographs, and/or a supporting diagram.

• Label the back (top surface) of the lift with appropriate markings.

• Subsequent lifts from the same impression must be numbered or otherwise labeled to distinguish which was first, second, etc.

• Case notes shall have some statement(s) addressing the quantity and description of impressions located, along with which ones were lifted. This may be aided with, but not replaced by, photographs.
7.6.3.5. **Adhesive and gelatin lift**

Adhesive lifting covers both gelatin and tape devices and permits lifting of some impressions when the ESDL is not optimal, available, or was unsuccessful. Adhesive lifters should only be used if the evidence cannot be transferred to the laboratory and after photography.

A gelatin lift can be used on porous or non-porous surfaces for lifting original residue deposit impressions that have been dusted with fingerprint powder, or impressions enhanced with some chemicals.

Tape devices, on the other hand, should only be used on non-porous surfaces and work best with impressions that have been dusted with fingerprint powder.

**Equipment**

- *Adhesive lifters* are generally tape of some kind. Adhesive lifters are generally purchased; however, fingerprint tape and other tapes can also be used.

- *Gelatin lifters* are commercially available in three colors: black, white, and clear.

- Clean fingerprint roller

**Method - Gelatin Lifter**

- Forensic scientists should take examination quality photographs of an impression prior to any lifting attempts.

- Choose the color of gelatin lift based on what will result in the best contrast between the target impression and the backing color. Black often works well for dusty shoe impressions, while an impression developed with black fingerprint powder may best appear on a white background.

- Remove the gelatin lift from the package and allow it to come to room temperature for 5-10 minutes prior to lifting an impression. This allows the gelatin to “relax” and will yield a more accurate size of the impressions.

- Cut the gelatin lifter to a size just larger than the target impression, if desired.

- Label the back of the lift to document the orientation, position, and location of the impression. This may also be done with mid-range photography that demonstrates the orientation of the impression in relationship to the surface it’s deposited on.

- Peel the protective transparent cover away from the gelatin layer.

- Place one edge of the lift on the substrate and slowly smooth down the rest of the lifter over the impression, taking care to
roll or press out any air bubbles. A roller may assist with this.

• After the lift has been smoothed over the entire surface of the impression, carefully remove it by the corners.

• Place it on a horizontal surface with the gelatin layer up and evaluate the results. The results should be evaluated in a darkened room with strong oblique light.

• Take examination quality photograph(s) of the lifted impression as soon as possible because the impression will fade over time. Unsuccessful lifts do not need to be photographed.

• To secure the lift for packaging, either cover with the protective plastic cover avoiding air bubbles, or tape it by the edges or corners (gelatin layer up) in the bottom of a flat cardboard box.

Method – Adhesive Lifters

• Take examination quality photographs of the impression prior to any lifting attempts.

• Place the adhesive lifter/tape at the edge of the impression and press or roll the tape across it, avoiding air bubbles. Repeat with overlapping tape lengths, if necessary.

• After the adhesive has been smoothed over the entire surface, carefully remove it from the corners.

• Place the adhesive on a clean fingerprint card, piece of paper, transparency sheet, or other similar product depending on which is likely to offer the best contrast for later imaging.

• Mark the lift to document the orientation, position, and location of the impression. This may also be done with mid-range photography that demonstrates the orientation of the impression in relationship to the surface it’s deposited on.

Cautions/Safety

• Adhesive lifts should not be attempted on non-porous substrates.

• Gelatin lifts will melt between 40° and 45° Celsius (104° and 113° Fahrenheit). Objects that have been exposed to the sun or these temperatures will need to be cooled before attempting to use a gelatin lifter.

• Lifts may be stored at room temperature, though storage in a refrigerator is advantageous.

• Attempts to lift dust prints on dirty backgrounds will cause both the dust print and dirty background to lift together. Subsequent lifts should be attempted to see if they would be successful.
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- Never slide the gelatin lift across the surface with the impression.
- If using a fingerprint roller to eliminate air bubbles in the gelatin lifter, do so gently to avoid shifting or excessive pressure that may damage the impression.
- Gelatin lifted impressions may fade and ultimately disappear over time.
- The rate of fading depends on storage temperature (cooler is better) and the material of the impression (e.g., silver fingerprint powders have lasted for several years without fading).

**Documentation**

- Mark and/or photograph the lift to allow for orientation.
- Label the back (top surface) of the lift with the appropriate descriptors.
- Subsequent lifts from the same impression must be numbered or otherwise labeled to distinguish which was first, second, etc.
- Case notes shall have some statement(s) addressing the quantity and description of impressions located, along with which ones were lifted. This may be aided with, but not replaced by photographs.

7.6.3.6. **Casting**

Casting is a technique that is used to collect three-dimensional impressions. Three-dimensional impressions are those that have a significant depth to them, in addition to the length and width of the impression. Commonly, they may be found in soil, sand, or snow. The detail within the impression may vary according to the substrate.

**Equipment**

- Dental stone or other forensic casting material (e.g., Traxtone, Crime-Cast™, etc.)
- Water
- Bucket or plastic bag
- Stir stick/spoon
- Tongue depressor
- Adjustable metal forms (approximately shoeprint size)
- Tray (plastic or metal)
- Snow Print Wax™
General Casting Method

The dental stone is mixed with water to a consistency approximating thin pancake batter. This mixture is then gently poured into the impression and allowed to harden. A tongue depressor can be held a few inches above the impression and the mixture poured along that and then into the impression, to direct the flow and reduce the chance of damaging detail in the impression. Also, sticks may be placed on top of the casting material to help support the cast after it has dried.

The required amount of dental stone will vary depending on the size of the impression to be cast. Therefore, variations are expected. One recommendation of volume/quantity is described below. Follow the manufacturer’s directions for other forensic casting materials.

Two (2) pounds of dental stone may be placed into an 8x12 inch Ziploc plastic bag. This amount will cast an average sized shoe impression. In preparation for use at crime scenes, numerous 2 lb. bags can be prepared and stored.

- Retrieve a two-pound bag, add about 10 ounces of water, and thoroughly mix in the closed bag. The mixture should have the consistency of thin pancake batter. If needed, add more water or dental stone to create the correct consistency.
- Metal forms may be placed around the impression to contain the casting mixture. These are less critical with the advent of dental stone and other forensic casting materials.
- Open the bag and with the bag at ground level, carefully pour the mixture into or next to the impressions, allowing it to gently flow into it. Fill the impression completely so that the mixture overflows out of the impression.
- When the cast is firm but still soft, scratch identifying marks on the exposed surface or write identifying marks with a permanent marker when the cast is dry.
- Allow the cast to dry for a minimum of twenty minutes in warm weather, longer in cold, wet conditions.
- Carefully lift the cast. Do not try to clean it; cleaning will occur in the laboratory.
- Package the cast in a large brown paper bag or cardboard box (not plastic) and allow to dry for an additional 48 hours.

Method for Underwater Impressions

Impressions that are underwater may still be cast. Do not attempt to drain
away any of the water as it is unnecessary and may disturb the impression. Place a metal casting form around the impression taking care not to distort/disturb it – the top of the metal form should be above the water line. Lightly sprinkle the dental stone over the underwater impression until covered by about an inch of the casting material. Then mix the dental stone mixture to a slightly thicker consistency than typical and carefully scoop the mixture onto the impression. Allow to set for at least 60 minutes.

Impressions that are shallow may be cast using the standard procedure except with a mixture that is slightly thicker – the mixture will displace the thin layer of water.

7.6.3.7. Method for Casting Impressions in Snow:

**DENTAL STONE**

- Using only the Dental Stone powder, sift a thin layer over the impression.
- Allow 1-2 minutes between each additional layer to allow for adequate hardening of the casting material.
- Repeat the above steps until there is no more moisture for the Dental Stone to absorb.
- Mix the remaining packet of Dental Stone with water and apply slowly to impression.

**SNOWPRINT WAX**

Impressions that are in snow should be sprayed with at least 3 to 4 layers of Snow Print Wax™ prior to casting, as it will preserve the detail. Allow to set for approximately 10 minutes. The wax shell is then cast with a slightly thicker-than-typical dental stone mixture that has been cooled.

- Be careful not to hold the can so close that the aerosol damages detail in the impression.
- Be sure that the entire impression is sealed with the Snow Print Wax or dental stone may seep through causing damage.
- Use cold water or some snow to help offset the exothermic reaction of mixing dental stone with water.
- Follow manufacturer recommendations for Snow Print Wax™.

**Cautions/Safety**

- Take examination quality photographs of the impression prior to casting.
- Do not clean out debris that is part of the impression or was present when the impression was made.
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- Use of fixatives is not recommended as they are not required to provide a protective layer to the cast (when dental stone or other forensic casting material is used), they rarely have any effect on preserving detail in the impression, and with improper use may obscure detail. If the forensic scientist chooses to use a fixative, they must note why it was necessary.

- A light dusting of a very fine powder or talc over the impression may be performed to prevent some of the soil from adhering to the cast, in an attempt to get a cleaner cast. Use of this technique is at the discretion of the forensic scientist, assuming the technique and powder used are fine enough to ensure that minute detail is not obscured.

- The reaction of dental stone with water is exothermic. This will have implications when casting impressions in snow (see Snow Impressions above).

- Once a dental stone mixture (or other forensic casting material) has hardened, it is not reversible. Use the mixture quickly after it is mixed, or it may harden in the mixing container. Thicker mixtures and warmer temperatures will cause hardening more quickly compared to thinner mixtures and colder temperatures.

Documentation

- Any impression(s) shall be photographed with examination quality photographs prior to casting.

- Label the back (top surface) of the cast with the appropriate markings using either a stick and scratching it when partially hardened or with a permanent marker when fully hardened.

- Case notes shall have some statement(s) addressing the quantity and description of impressions located along with which ones were cast. This may be aided with, but not replaced by, photographs.
7.6.3.8. **Mikrosil Casting**

Mikrosil™, a silicone rubber casting material, is a two-part silicone type material available in brown, white, and black colors. Mikrosil™ is useful to the forensic scientist to collect and preserve three-dimensional toolmark evidence from a scene that can’t be reproduced in photographs alone. Additionally, Mikrosil™ can be used to lift latent prints developed on irregular surfaces.

**Equipment**

- A Mikrosil™ package contains one large tube of silicone rubber casting material and a second smaller tube of silicone hardener (catalyst).
- Mikrosil™ Base (White, Brown, Black, and Grey)
- Mikrosil™ Hardener
- Wooden tongue depressor or metal spatula
- Mixing surface (fingerprint lifting card will do)
- String tags (optional)

**Method**

Mikrosil™ is available in four colors: brown, white, grey, and black. The brown color is recommended for the reproduction of toolmark impressions. White Mikrosil™ is recommended for lifting latent print impressions that have been developed with black powder, and black Mikrosil™ is recommended for lifting latent print impressions that have been developed with a light color powder.

It is advised that the forensic scientist prepare a label for the Mikrosil™ cast since it is nearly impossible to write on the silicone rubber.

Remove any debris that may be in the impression.

**Prepare the Mikrosil™**

1. From the large tube, place a line of Mikrosil™ enough to cover the impression onto the mixing surface.
2. From the smaller tube, place a line of hardener next to the line of Mikrosil™; both lines should be the same approximate length.
3. The proper ratio of hardener to Mikrosil™ is 1:1.5, but more catalyst will reduce the working time and less catalyst will increase the setting time.
4. Thoroughly mix the two lines of Mikrosil™ together using a tongue depressor or metal spatula. This should take 30 to 60 seconds.

**Application of Mikrosil™**

1. Remove the mixed Mikrosil™ from the card and apply to the surface taking care to work the casting material into or on the impression, but not allowing the mixing tool to come in contact with the impression.
2. Place the prepared label in the Mikrosil™ and allow the cast to set. A
wooden tongue depressor may be pressed into the Mikrosil™ cast before it sets when a label or tag is not practical.

3. The setting time will be approximately 5 to 8 minutes in 68-degree farenheight temperatures and 12 to 15 minutes in below-freezing temperatures with a standard ratio of Mikrosil™ and catalyst.

4. It is recommended that a photograph be taken of the Mikrosil™ cast on the area prior to its removal to aid in the orientation of the cast.

5. Gently peel the Mikrosil™ from the impression after it has setup.

Documentation
 Label and orient the lift with the appropriate labels.

Packaging
 A newly recovered cast may become stuck to another recovered Mikrosil™ cast if they are packaged together. For this reason, the cast should be packaged separately. Either in a small box or envelope so that multiple casts do not make contact and become stuck.

Alternatives to Mikrosil™
 Forensic Sil, Accutrans®, or other similar based products manufactured by different companies can be used. These products are mixed and dispensed as follows:

Equipment
 These casting materials come in dual-stage tubes, which include both casting material and a special catalyst.

Method
 Load a tube into the dispensing gun. Squeeze the trigger to mix casting material and catalyst on demand through a special mixing tip. No mixing is required, and you only use as much casting material as needed. Leave the applicator tip on the tube once finished. These products won’t dry in the tube. When you are ready to make another cast, simply replace the used tip with a fresh one.

Documentation
 See casting with Mikrosil documentation.

Packaging
 A newly recovered cast may become stuck to another recovered cast if they are packaged together. For this reason, the cast should be packaged separately. Either in a small box, envelope or packaged in such a manner that multiple casts do not come into contact with each other.
7.6.3.9. **Tire Exemplars**

The exemplar provides a record of the characteristics present on an item of evidence at a given time.

The impression a tire leaves will be slightly different depending on whether or not it is under load. If exemplars are being made for purposes other than elimination, they should include the full circumference of the tire when under load. Typically, this is between six and eight feet in length. When the actual vehicle is unavailable, the tire(s) may be mounted on a similar vehicle. The tread elements will not change significantly with slight air pressure or load variations.

**Elimination Exemplars**

Partial exemplars may be collected for purposes of elimination of vehicle tires or for documentation of an obvious exclusion based on tread design differences by a Footwear and Tire Tread examiner.

**Imaging**

An image of the tire tread is sufficient. A scale shall be included in the photograph of the tire tread. Information about the tire and vehicle should be recorded.

**White Adhesive Lift**

Using a large adhesive lift, the residual material on a tire can be lifted off, providing a good representation of a section of the tread design. Information about the tire and vehicle should be recorded.

**Full Tire Exemplars**

When it is determined that the collection of tire exemplars is to be performed at the time of the vehicle process, the forensic scientist should first capture photographs that are easily available/collectable. The forensic scientist should then seek the assistance of a Footwear/Tire Tread examiner. It may be beneficial to provide the Footwear/Tire Tread examiner with images of the impressions and/or vehicle tires ahead of time. In some cases, the collection of full exemplars may be deemed unnecessary. This shall be determined by the Footwear/Tire Tread examiner.

Note: If comparison of individual characteristics will be requested, the collection of the tires is required even when tire exemplars have been collected. The conclusions of a subsequent comparison may be limited if the tires are not available for examination.
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Equipment
Broom, Kraft paper, duct tape, work gloves, scissor/utility knife, chart board, wet media film, tape/chalk, sharpie or other markers, cloth measuring tape, either petroleum jelly or silicone oil, magnetic fingerprint powder, magnetic fingerprint powder brush, clear lacquer spray or printer’s ink, clear plastic sheeting.

Method (VEHICLE/TIRE PREPARATION)
Before using any of the techniques listed below, the following steps shall be performed:

• Find an adequate surface to collect the tire exemplars. A smooth area of asphalt or concrete is sufficient and should be swept clean. The size of the area needed will depend upon the vehicle and tires, but will need to be at least long enough to permit one full tire rotation with enough extra room to bring the vehicle to rest from the exemplar sample.
• Roll out enough paper to keep the tires from collecting debris while moving. Secure the paper with tape.
• Clean the tires by rubbing lightly while wearing work gloves. Cleaning should be to remove surface debris only. Rock-holds, etc. should be left in place.
• Measure the circumference of the tire(s) of interest using a cloth measuring tape.

• Measure the wheelbase of the vehicle.
• Mark the tire with tape at 5 to 6 points equidistant around the tire and label alphabetically.

• The number of points may vary at the examiner’s discretion depending on the size and design of the tire. Document the location of the labels on the tire with enough detail (e.g. close-up photography) that the labels can be re-created and/or replaced at a later date.

• Record the information from the tire side wall.
  o The make/model/year of the vehicle
  o Make/model of the tire
  o DOT number of the tire
  o P-metric tire size designation
  o Mold numbers when possible

Making Exemplars Using the Petroleum Jelly/Silicone Oil, Fingerprint Powder on Wet Media Film Method

It is important to use only a small amount of petroleum jelly/silicone oil or detail will be lost or obscured. This method provides good detail and a transparent background that facilitates the comparison process.

• Tape sections of chart board together to achieve a length slightly longer than the tire circumference, typically 4-6 extra inches. If using wet media film, secure a length of film on top of one of the lengths of chart board.

• Using gloved hands, rub a very small amount of petroleum jelly or silicone oil to coat one or both of your hands.

• Then thoroughly rub the tread surface of the tire so an even, thin coating is applied to the full circumference.

• Place an end of chart board (with film) just in front of the tire. Drive the vehicle in a continuous motion over the chart board, depositing an impression on the film. As the tire
rolls, mark the locations on the exemplar where the A-F labels on the tire correspond.

• Develop the impression by powdering it with magnetic fingerprint powder.
• Spray 3-4 coats of a fixative, such as a clear lacquer to preserve the exemplar and allow to dry as per manufacturer’s instructions prior to handling or packaging. The impression will be damaged if it is touched before dry.
• Mark the exemplar with pertinent case information that includes the location and orientation of the tire.
• Two exemplars from each tire are recommended. The exemplars should be off set so that they do not end in the same location.

Making the Tire Exemplars Using Black Printer’s Ink

• Tape sections of the chart board together to achieve a length slightly longer than the tire circumference, typically 4-6 extra inches.
• Repeat this step.
• Apply a thin layer of printer’s ink to one of the sets of chart board.
• This will be the ink pad.
• Roll out plastic sheeting to achieve a length slightly longer than the tire circumference, typically 4-6 extra inches.
• Tape clear plastic sheeting to the other set of chart board.
• Line the chart board sets up so that the tire is inked and then rolls over the plastic sheeting.
• Drive the vehicle in a continuous motion over the chart board, depositing an impression on the film. As the tire rolls, mark the locations on the exemplar where the A-F labels on the tire correspond.
• Mark the exemplar with pertinent case information that includes the location and orientation of the tire.
• Two exemplars from each tire are recommended. The exemplars should be off set so that they do not end in the same location.
• Allow the ink to dry. This may take overnight or longer.

Documentation

The following information shall be recorded in the case notes regarding exemplars:

• Method of exemplar collection
• The number of exemplars collected from each tire
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- Tire sidewall information (as outlined above in VEHICLE/TIRE PREPARATION)
- Vehicle information (make, model, year, VIN)

The following information shall be recorded on each exemplar collected: case #, date of collection, initials of the examiner, location of the tire on the vehicle (e.g. front right, rear left, etc.), direction of travel, where on the exemplar the specific tire location designated A-F corresponds, direction towards front of vehicle, indication of the outside and inside edge of the impression (as the tire is mounted on the vehicle).

**Storage**
Once the exemplars are completely dry, they can be rolled up and packaged.
7.6.4. **Trace Evidence**

Trace evidence is a generic term for small often microscopic material. Small items such as fibers, hairs, broken glass fragments, paint fragments, and assorted microscopic debris may be left by a person or picked up from contact with the environment or another person. Trace samples may ultimately aid in determining origin to link to a person or place.

The analyst will typically use the particle picking or tape lifting technique. The choice of collection method may depend upon the evidence type of substrate and the need to determine exact location of evidence, among other variables.

Ada County Forensic Lab does not conduct trace evidence analysis; however, processing, collection and documentation may still be performed at a crime scene by forensic scientists.

**Collection of Trace Evidence**

There are a variety of methods that may be used to collect trace evidence. The choice of collection method will depend upon variables such as the evidence type, the substrate, and the need to determine the exact location of the evidence.

**Particle Pick**

Particle picking is the recommended technique when visible trace evidence is to be collected. This is the only method that allows the forensic scientist to determine the exact location a specific piece of trace evidence was recovered from.

**Equipment**

Particle picking may be performed without equipment, using only the forensic scientist’s gloved fingers, or with the aid of Post-it notes, tweezers, forceps, or other tools.

**Method**

- Visually examine the evidence with the unaided eye and/or with the aid of a magnifying lens.
- When trace evidence is located, use either gloved fingers or a tool (e.g., forceps, tweezers, etc.) to collect the trace evidence and place on the adhesive of a Post-It note and then fold over, in a paper fold, glassine envelope, or other appropriate container and label.

**Cautions/Safety**

- There may be trace evidence that is not visible. Therefore, additional collection techniques may need to be employed to ensure collection of non-visible trace evidence.
- Use of tools, such as forceps and tweezers, may cause damage
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to the trace evidence.
- Clean any collection tools thoroughly between samples to prevent cross-contamination.
- Avoid serrated tools as they may be more difficult to clean thoroughly.

**Adhesive/Tape Lifts**

Adhesive/tape lifts are used to collect trace evidence from a variety of surfaces, such as vehicle seats and clothing items. This method of collection is not recommended for substrates that will strongly adhere to the tape lift adhesive (e.g. paper products, cardboard, etc.). This method is useful when collecting trace evidence that is not visible or apparent to the unaided eye. The technique collects trace evidence from an area defined by the forensic scientist. Determining the exact location of a specific piece of trace evidence may not be possible.

**Equipment**

Adhesive/tape lifters, clear and colorless (e.g., adhesive lift sheets whole or cut to smaller sizes), scotch tape, fingerprint lifting tape, mailing tape.

Plastic sheets, clear and colorless (e.g., transparency film, sheet protectors), may be pre-printed with a grid to aid in later examination.

**Method**

- Expose the adhesive layer of the adhesive/tape lift. Do this just prior to use to avoid contamination.
- Repeatedly pat the adhesive/tape lift in the target area, being careful not to overload the adhesive surface.
- Place the lift onto a clear plastic sheet to protect the collected trace evidence from contamination, damage, or loss.
- Repeat the process until the area defined is finished.
- Package in an envelope or other container to prevent contamination, damage, or loss.
7.6.4.1. **Trace Standards**

7.6.4.1.1. **Hair Standards**
Microscopic hair comparisons may be performed on human head and pubic hairs. Hairs from the same body region of one person are known to have a variation in characteristics; therefore, it is important to obtain enough hairs to adequately represent the range of all characteristics present.

**Head Hair Standards**
A complete head hair standard consists of at least 24 to greater than 50 hairs collected from five (5) different areas of the scalp: center, front, back, and both sides. Hairs that are cut are not an appropriate standard.

- Collect loose or shedding hairs. Use a clean, new comb to vigorously comb through the head hair of the subject. Be sure to collect hairs from the center, front, back, and both sides. Do this over a clean piece of paper or envelope to collect hairs that are dislodged.
- Pull hairs from various areas of the scalp by grasping a hair(s) firmly near the root and pulling quickly. Place collected hairs into the same package as the loose/shed hairs.
- Repeat these steps until at least 24 hairs are collected.
- Secure the paper fold or seal the envelope to prevent contamination, damage, or loss of the hairs.
- Label the outside of the package with the individual's name and the body region it was collected from.

**Pubic Hair Standards**
A complete pubic hair standard consists of at least 24 to greater than 50 hairs collected from different areas of the pubic region. Hairs that are cut are not an appropriate standard.

- Collect loose or shedding hairs. Use a clean, new comb to vigorously comb through the pubic hair of the subject. Be sure to collect hairs from different areas around the pubic region. Do this over or near a clean piece of paper or envelope to collect hairs that are dislodged.
- Pull hairs from various areas of the pubic region by grasping a hair(s) firmly near the root and pulling quickly. Place collected hairs into the same package as the loose/shed hairs.
- Repeat these steps until at least 24 hairs are collected.
- Secure the paper fold or seal the envelope to prevent contamination, damage, or loss of the hairs.
- Label the outside of the package with the individual's name and the body region it was collected from.
Secondary Hair Standards

A secondary standard is not obtained from an individual directly, but from an object or location where the individual is believed or known to have deposited hair (e.g., a hairbrush). Necessity should be the only reason to obtain secondary hair standards vs. pulled/combed hair standards (e.g., missing person).

Secondary hair standards may be obtained if it can be demonstrated or documented that the hair collected from the object/location is unlikely to include hair(s) from other individuals.

Cautions/Safety

Do not package hair standards from different sources or areas in the same envelope or package hair standards with hair evidence. This could allow cross contamination to occur.

Securely seal all possible openings in packaging, including seams and air holes.

Avoid stretching or breaking the hairs when pulling them.

7.6.4.2. Fiber Standards

Fiber comparisons are performed on both natural and synthetic fibers. A potential fiber source may have one or more different kinds and colors of fibers that are present. The differences may only be apparent using microscopic or instrumental techniques; therefore, it is important to obtain a fiber standard that adequately represents all of the fiber types present in the potential source.

A fiber sample that is being excised from a potential source must be large enough to capture the variation of fiber types present. Differences in the color or texture of a fabric, carpet, or other source should alert the forensic scientist that different fiber types might be present so a standard must be collected from each discernible area.

Moveable Objects

- If the potential source of a fiber transfer can be packaged and transported to the laboratory with ease, then it should be submitted in its entirety (e.g., clothing items, throw rugs, etc.)
- Package moveable items in clean, and previously unused, packages such as envelopes, paper, and plastic bags.
- Securely seal and label the package with a description of
where the standard came from and other appropriate markings.

**Immoveable Objects**
If the potential fiber source is from a large object or one not easily transported, such as car upholstery or carpeting from a dwelling or vehicle, use the following method to collect a fiber standard.

- Cut a representative sample from various areas of the object. Be sure the cut is deep enough that the backing material or substrate is also collected.
- If the object appears uniform, only one sample needs to be collected.
- Collect samples that are visually different (e.g., different colored areas, faded areas due to sunlight, worn sections, etc.)
- A sample size of approximately 1 X 1 inch is fine unless variations are visible. Differences warrant a larger size cutting. If the source appears uniform, a smaller size cutting may be acceptable.
- Package in an envelope, paper bag, plastic bag, or other container.
- Securely seal and label the package with a description of where the sample came from and other appropriate markings.

**Cautions/Safety**
- Do not pull or tape lift fiber standards.
- Securely seal all possible openings in packaging, including seams and air holes.
- Do not allow potential fiber sources to come into contact with fiber evidence samples. Cross contamination may occur.

7.6.4.3. **Paint Evidence**
Paint comparisons are performed on a variety of paint types including vehicle paints, architectural paints, spray paints, cosmetic lacquers, etc. A potential paint source may have one or more different kinds and colors of paint that are present (e.g., vehicles). The differences may only be apparent using microscopic or instrumental techniques; therefore, it is important to obtain paint standards that adequately represent all the paint types present on a potential source.

The forensic scientist should consider the possibility of a physical match of paint chips to the source when determining
how to collect paint standards.

**Moveable Objects**

If the potential source of a paint transfer can be packaged and transported to the laboratory with ease, then it should be submitted in its entirety (e.g., painted crowbar, bicycle, etc.).

- Package moveable items in clean, and previously unused, packages such as envelopes, paper, paper, and plastic bags.
- Securely seal and label the package with a description of where the standard came from, along with any other labeling requirements.

**Immoveable Objects**

If the potential paint source is from a large object or one not easily transported, such as a vehicle, use the following method to collect a paint standard:

- Locate the area of damage, if applicable (e.g., damaged vehicles).
- Collect from an area as close to, but not within the point of damage. If a physical match examination is deemed appropriate, collect all the damaged body panels rather than attempting to remove paint standards.
- Use a clean razor blade, scalpel, or appropriate tool to gently pry, carve, or chip the paint from the surface down to the foundation or substrate. If possible, do not remove the paint by scraping as all paint layers may not be represented and/or the layer structure may be destroyed.
- Collect about a nickel-sized combined amount of paint from a particular damaged area, when possible.
- Place the paint evidence into a paper fold or small paper envelope, carefully sealing to prevent loss.
- Securely seal and label the package(s) with a description of where the sample came from, along with any other labeling requirements.
- Continue to collect paint from each damaged area in the same manner, even if the object appears uniformly painted. Also collect samples that are visually different. Package and label each area separately.
Cautions/Safety

- Substantial variations in thickness and layer sequences over short distances can exist across a painted surface. This is particularly true in architectural paint and for vehicle paint where curves, corners, and edges are often impact points and may have been subjected to previous damage, sanding, or over-painting. Known paint samples should be collected from these areas when recognized.

- When contact between two painted surfaces is indicated, the possibility of cross-transfers must be considered. Collect both objects or paint standards from both surfaces.

- Securely seal all possible openings in packaging to include seams, corners, and air holes.

- Do not allow potential paint sources to come into contact with paint evidence samples. Cross contamination may occur.

7.6.4.4. Glass

Glass comparisons are performed on window glass, vehicle glass, object glass, and other glass types.

A critical factor in comparing glass evidence to a potential glass source is whether the characteristics of the evidence sample fall within the range of variation present in the source; therefore, it is important to obtain a sufficient number of glass samples to adequately represent the range of all characteristics present.

The forensic scientist should consider the probative value of a physical match of glass to the source and the determination of the direction of force.

Method

Collect as much of the original glass as possible.

Packaging

- Package each standard collected separately in a cardboard box or other rigid container (do not use glass containers). Secure the pieces to reduce the likelihood of further breakage or damage to the fractured edges. Small glass particles may be packaged in folded Post-it notes, paper folds, envelopes, paper bags, etc. Then small packaged particles can be placed in a padded envelope to prevent damage and/or injury.

- Securely seal and label the package with a description of where the standard came from.
Cautions/Safety
Broken glass edges are extremely sharp. Handle with caution. Use personal protective equipment or tools to reduce the risk of being cut.

Other Types of Trace Evidence
Other types of trace evidence to consider when processing a scene may include but is not limited to soil, foliage, metal fragments, and wood fragments.
7.6.5. **Firearms/Toolmark Evidence**

Firearms evidence can include firearms, ammunition, discharged cartridge cases, projectiles/fragments, bullet impact marks, and gunpowder residue.

**Collection and Packaging of Firearms Evidence**

The primary concern during the collection and packaging of a firearm is safety. **ALWAYS HANDLE A FIREARM AS IF IT IS LOADED AND KEEP IT POINTED IN A SAFE DIRECTION.**

Preservation of evidence that may be present on the firearm (e.g., blood, trace evidence, latent prints) shall be assessed prior to packaging the evidence.

When collecting a firearm, pick it up by the textured surface on the grips or by the edges of the trigger guard. Never move a firearm by inserting an object inside the barrel or trigger guard. This is unsafe and can damage potential evidence.

Transitory evidence (e.g., hairs, fibers) should be documented at the scene and collected prior to packaging the firearm. If dried flaking blood is present, the firearm should be wrapped in paper (prior to any additional packaging, such as a gun box) to contain any dislodged evidence. When a firearm is collected from the scene, the forensic scientist should consider whether swabs should be collected for DNA preservation purposes before any other additional testing analysis.

The final step is securing the firearm in a box, with a tag, or package in paper.

**Revolver**

The following guidelines should be used when handling a revolver:

Photograph the firearm prior to handling or collecting it. If the firearm was moved prior to your arrival, also note the reported original position and the name of the person who reportedly moved the firearm.

Document the position of the hammer. If the revolver is cocked (hammer back), carefully let the hammer down by manipulating the trigger while holding the hammer spur.

Mark the position of the cylinder on both sides of the top strap prior to opening the cylinder.

Document the direction of rotation of the cylinder (clockwise or counterclockwise).

Record the number of chambers and document the brand and condition (i.e., fired, unfired) of the ammunition in each chamber (refer to the following example).
After documentation cartridges can be packaged in paper evidence packaging. Fired cartridge casing should be packaged in a way that prevents any alteration or obliteration of microscopic markings. When the forensic scientist deems it necessary, cartridges/cartridge cases can be individually removed from the revolver and packaged separately in an envelope or paper bag to prevent alteration or obliteration of microscopic markings along with the documentation previously.

**Semi-automatic Pistols, Rifles, and Shotguns**

The following guidelines should be used when handling a gun:

- Photograph the firearm prior to handling or collecting it. If the firearm was moved prior to your arrival, also note the alleged original position and the name of the person who reportedly moved the firearm in the case notes.

- Document the position of the hammer. If the firearm is cocked (hammer back), carefully let the hammer down by manipulating the trigger while holding the hammer spur. Document the position of any manual safety devices and/or de-cocking levers.

- Carefully disengage the magazine and remove it from the firearm. Package the magazine in a paper bag, envelope, or zip tie in gun box.

- Open the action and visually check the chamber for a fired or unfired cartridge/shotshell. Remove any cartridge/shotshell, document the brand and condition (i.e. fired, unfired) and package it in an envelope or paper bag to prevent alteration or obliteration of microscopic markings.
7.6.5.1. **Trajectory Processing**

Trajectory Processing includes the identification and processing of trajectory evidence and the collection of measurements for reconstruction purposes. Observations of impact sites can provide information about the possible projectile, the type of firearm involved, intermediate objects in the path of the projectile, direction of travel (entrance or exit) and other details. In certain circumstances, the trajectory of the projectile may be determined and in turn, the possible positions from where a shot originated.

Identifying the possible position of the source (shooter[s]) reduces the sites to search for evidence and may provide investigative leads and supplemental information. Shooting incidents are dynamic and varied, as is the evidence produced during such an event. The methods outlined below demonstrate techniques that can used to process these scenes.

**Equipment**

The following items are suggested for trajectory, but is not a complete list: cameras, tripod, trajectory rods, centering cones, rubber O-rings, angle finder (digital or inclinometer), plumb bob, string, protractor, laser, mirrors and calculator.

**Cautions/Safety**

The lasers used for these purposes can be intense and exposure to the eyes can cause damage. Never look directly into the beam and always alert others in the area before the laser is turned on.

Lead is typically present on or in projectiles associated with shooting incidents and can be found in holes and impacts created by these projectiles. Exposure to lead can be hazardous.

**Method**

These are the following guidelines for documenting and processing trajectory evidence:

- Bullet hole/defect shall be photographed prior to any attempt to determine the trajectory (overall, intermediate and close-up, without and with scales and evidence markers).

- If multiple defects are present, unique identifiers shall be used in the documentation to differentiate between the holes that are observed.

- Document the location of the bullet hole in your notes (sketch, measurements, diagram).

- All trajectory measurements should only be collected when
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deemed to have an appropriate entrance and exit. The reliability can increase/decrease depending on the substrate and number of bullet holes associated with the trajectory. This is up to the Forensic Scientist’s discretion on the reliability of the trajectory and will proceed with data collection based on this knowledge.

Measurements consist of horizontal and vertical angles, and locations of the bullet holes.

Determine the direction of bullet travel, if possible (entrance vs. exit).

Document the initial impact bullet hole’s size. This can be done on scene by measuring or via photograph with a scale.

If applicable, collect, and document trace evidence. This type of evidence may have been transferred from the projectile itself or may indicate that an intervening object may have been involved.

Observations such as beveling, paint fractures, pinch point, lead-in marks, etc. can be included in the documentation.

The choice of the measurement tool that is used (laser angle finder, inclinometer, etc.) to determine the trajectory is up to the forensic scientist.

Using the trajectory kit:

- After documenting the primary bullet hole’s size, carefully place a trajectory rod through the entrance hole and exit hole.
  - If applicable, place a cone shaped centering cone on the rod and gently guide the tapered end into the entrance hole and use a rubber O-ring to hold the guide in place. Use a second centering cone for the exit hole.
  - Forensic scientists should consider the stability of the rod when determining trajectories. It’s up to the examiner to determine stability. A stable rod should be used for trajectory determinations.

- Determining Vertical Angle—Place the angle finder along the top of the trajectory rod and record the measurement of the vertical impact angle relative to the horizontal plane. Alternatively, a laser angle finder or protractor can be used to determine the impact angle. With the protractor in a vertical position, place the center of the flat edge flush with the center of the bullet hole. From a 90° angle (side-view), determine the vertical impact angle. It is important to document whether the measured angle is in relationship to the target surface or the horizontal plane. This can be documented by a description and/or a sketch.
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- Determining Horizontal Angle—With the protractor in a horizontal position, place the center of the flat edge flush with the center of the bullet hole. Alternatively, a laser angle finder or protractor can be used to determine the impact angle. From a 90° angle (top-view), determine the horizontal impact angle. It is important to document whether the measured angle is in relationship to the target surface or the vertical plane. This can be documented by a description and/or a sketch.

- A laser or string can be attached to the end of the trajectory rod to project/extend the trajectory in either direction.

For demonstrative purposes, the trajectory can be photographed in conjunction with trajectory rods, strings and/or a laser and photographic fog. The forensic scientist shall be cognizant of the effect that the placement of the camera in relationship to the trajectory will have on the resulting photograph (the trajectory can appear skewed due to the angle and level of the camera).

When trajectory processing is complete and if circumstances warrant, collect the wall surround a bullet hole, ensure that there is adequate room that allows for later processing by firearm examinations.

**Reporting**

The forensic scientist shall report the results of any trajectory determinations that were made. For the purpose of the lab report, it is recommended the forensic scientist use directional descriptors with indications of upward or downward, right or left, for ease of understanding the determined trajectories. Whenever possible, reported origins of projectiles should be given in context with the scene rather than just numbers or directions.

If numerical values were collected, those values will be stored for a reconstruction report. If requested, the forensic scientist will report this information that was collected and retained for future use. If a forensic scientist chooses to report numbers for the angle determinations, a footnote or statement shall be added to reflect a ±5° variance as reflected in current readings and teaching within the field. Any reported numbers shall be clearly reported as approximate.

Modifiers such as “slightly” or “acutely” can also be used. Additional descriptions of the relative location of the impacts and the terminus of the bullet are also helpful examples of supporting documentation that should be included in the report.
7.6.5.2. **Bullet impact presumptive tests**

When a fired bullet impacts a surface at high energy the surface of the bullet can transfer onto the surface impacted. Bullets are primarily composed of lead or copper/brass jacketed lead. Sodium Rhodizonate (NaRho) can detect the presence of lead and Dithiooxamide (DTO) can detect the presence of copper.

Dithiooxamide (DTO) will not destroy (dissolve) lead if it is present at the impact site. Conversely, acetic acid, present in Sodium Rhodizonate, can dissolve and remove copper. Therefore, tests to detect copper must be performed prior to tests for lead. If there is a question of entrance vs. exit, both sides of a hole should be tested.

7.6.5.2.1. **Dithiooxamide (DTO) for copper**

Dithiooxamide (DTO) can detect the presence of copper. When a copper jacketed projectile impacts a surface, trace residual copper may be left behind. DTO will produce a green-grey color in the presence of copper.

**Reagents**

- **Dithiooxamide Solution**
  0.2% solution dithiooxamide in ethanol (w/v):
  (Mix 0.2 g DTO in enough ethanol to make 100 mL)

- **Ammonium Hydroxide solution**
  2 parts concentrated ammonium hydroxide to 5 parts water.
  (Mix 20 mL concentrated ammonium hydroxide plus 50ml deionized water.)

**Method**

- Place 3 drops of ammonium hydroxide solution on swab or filter paper.
- Place the swab or filter paper over the area to be tested.
- Remove the filter paper and place 3 drops of dithiooxamide solution to the tested area of the swab or filter paper.

**Interpretation**

- Positive: A positive reaction will show a dark greenish-gray color.
- Negative: No color change should be observed.

**Controls**

- Positive: Wet a swab or filter paper with the ammonium hydroxide solution and run it on a known piece of copper. Then add the DTO solution.
- Negative: Wet a swab or filter paper with the ammonium hydroxide solution. Then add the DTO solution.
  Working solutions checked at the time of use.
Inconclusive: A positive reaction that occurs prior to the addition of ammonium hydroxide and dithiooxamide solution, development of a different color, or other circumstance that leads the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination.

7.6.5.2.2. **Sodium Rhodizonate (NaRho) for lead**

Sodium Rhodizonate (NaRho) allows lead-containing residues in bullet wipe and/or projectile impact sites to be made visible by a simple color-complexing reaction. Sodium Rhodizonate (NaRho) will produce a pink color after the addition of Sodium Rhodizonate followed by a violet color after the addition of hydrochloric acid.

**Reagents**
- **Saturated solution**
  - Saturate water with sodium rhodizonate (.27g in 200ml water used)
- **Hydrochloric Acid Solution**
  - Hydrochloric acid 5 mL
  - Deionized water 95 mL
- **Buffer Solution**
  - Sodium bitartrate 1.9 grams
  - Tartaric acid 1.5 grams
  - Deionized water 100 mL
  - *This may require heat and agitation.
- **Acetic Acid Solution**
  - Glacial acetic acid 15 mL
  - Deionized water 85 mL

**Method**
- Dampen a swab or filter paper with acetic acid solution.
- Swab the area of interest or use filter paper.
- Apply 1-2 drops of sodium rhodizonate solution to the sample.
- Apply buffer solution to the sample.
- Apply hydrochloric acid solution to the sample.

**Interpretation**
- **Positive:** A positive reaction will show a pink color after the addition of sodium rhodizonate and a violet or purple color after the addition of the hydrochloric acid. Results can fade quickly.
- **Negative:** No color change should be observed
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Inconclusive: A positive reaction that occurs prior to the addition of hydrochloric acid solution, development of a different color, or other circumstance that leads the forensic scientist to be cautious about a positive or negative interpretation. Document the observation(s) that led to an inconclusive determination.

Controls

Positive: Wet a swab or filter paper with acetic acid and swab (or use filter paper) a piece of lead with it, and then apply solutions as listed above.

Negative: Wet a swab or filter paper with acetic acid and apply other solutions listed above.

Working solutions will be checked at the time of use
7.6.5.3. **Toolmark Evidence**

A toolmark is any impression, scratch, gouge, cut, or abrasion made when a tool is brought into contact with an object, leaving a mark.

**Photography**

Overall, mid-range and close-up (examination quality) photographs shall be taken of the toolmark. Close-ups shall be taken with a scale.

For toolmark photographs, these photographs should be uploaded to RMS/LIMS system for future use.

**Measurements**

Measurements should be taken to document the toolmark in relationship to the ground and/or other fixed objects.

**Casting/Collection**

If an item cannot be submitted for toolmark examination, a cast/photograph may be taken. Casts using a flexible casting material such as Mikrosil should be utilized. Refer to Mikrosil casting procedure.
7.6.6. **Latent Friction Ridge Evidence**

When the friction ridge skin area of the palmar or plantar regions of the body are touched to a receiving surface, a reproduction of the ridge design from that friction ridge skin area may be left behind on that surface.

**Latent Print Recovery**

When processing evidence consideration shall be given for all types of physical evidence and the order in which the evidence should be processed. The most common forms of latent print processing in the field are powder processing (dusting) and use of cyanoacrylate (CA). Other processing methods may be used depending upon the circumstance of a particular case. A latent print discipline expert shall be consulted prior to chemical processing.

In most circumstances, smaller items of evidence should be collected for processing in the lab. Field processing should be limited to large items that cannot be easily transported and fixed structures.

All areas processed will be assessed for the presence of visible friction ridge detail. Areas of ridge detail deemed to be suitable for collection will be marked with a unique identifier. The unique identifier must be such that the latent can be properly re-located in the scene and matched back to any lift cards collected or photographs.

Latent friction ridge evidence requiring chemical enhancement of suspected blood should utilize chemicals found in the presumptive blood test/enhancement section. It is ideal that evidence be collected and taken to the lab, but for items not capable of being transported to the lab, on-scene processing by an authorized forensic scientist should be performed. The forensic scientist should utilize one of the following chemicals for blood enhancement: LCV and/or Amido Black. Methods and other information can be found in the biology section under presumptive blood tests/enhancement section of this analytical method.

7.6.6.1. **Physical/Powder Development**

**Equipment**

- Gloves
- Latent print Powders
- Light source
- Camera equipment
- Nylon/fiberglass brush and/or magnetic wand
- Fingerprint tape
- Glossy fingerprint cards
Method
Use of an unused disposable brush and unused secondary container of powder ensures the risk of contamination is kept to a minimum if DNA is a concern. If there are no DNA concerns, re-hydration of the print can be considered. This is accomplished by lightly huffing on the area with the print, waiting for the moisture to dissipate and then applying additional powder.

Multiple prints in close proximity may be collected on the same lift. In some instances, one may need to perform multiple lifts (re-powdering of the same print to capture the best quality latent lift.)

Once lifted, the tape will be affixed to either a lift card of appropriate color (i.e., white cards for black and bi-chromatic powder) or a transparency cover. Lift cards will be marked with appropriate identifying information such that the card can be matched back to the item of evidence and/or the scene location.

If the forensic scientist believes it is unlikely that the impression can be lifted, then the forensic scientist should make all attempts to photograph, collect, and package the evidence with the area of ridge detail.

Fiberglass/Nylon Powder brushes
Only the ends of the brush bristles should be coated with the powder, and the brush should be gently tapped several times to remove all but a minimum amount.

With the brush handle in a nearly perpendicular position to the surface the bristle ends are lightly and delicately moved over the surface. Discoloration of the latent print residue will usually appear immediately. With a nylon/fiberglass brush and a proper amount of powder, the impression will develop in density with each light pass until no further development can be observed.

7.6.6.2. Magnetic Powder (Brushless)
Magnetic powder must be applied with a magnetic applicator.

Surface areas examined generally must be processed more slowly with magnetic powders. Great care must be exercised to prevent actual contact between the end of the wand and the surface.

Disposable, sterile magnetic wands may be employed (for DNA/contamination purposes), or wands may be retained and reused as long as they are decontaminated with a 10% bleach solution between scenes or sooner as applicable.
Cyanoacrylate ester

Cyanoacrylate (CA, i.e., superglue) fuming has been shown to be an effective means of latent print development on non-porous and some semi-porous surfaces (e.g., plastic, carbon paper, metals, glass, tapes, wood, rubber, and rock). Cyanoacrylate ester fumes are monomers that polymerize on latent print residue and create a more stable impression.

Equipment

CA at crime scenes is used in various forms: a commercial kit or packs, a fume wand or liquid CA in combination with a heat source.

Methods (Liquid, HotShot, CA Pad/Pack)

Liquid CA with heat source

Liquid glue is placed in a disposable container (aluminum foil works well), which is then placed over a heat source in the vehicle, processing area, or fuming chamber resulting in the production of fumes. Heating may be accomplished with a coffee cup warmer or a light fixture assembly (60-watt bulb). DO NOT USE A HOT PLATE OR DIRECT FLAME. Once the test print shows sufficient development, ventilate area to evacuate all fumes.

HotShot™

- These kits work rapidly and produce copious amounts of cyanoacrylate fumes. Each HotShot™ contains the plastic container, an Activator Packet, HotShot™ drum and a vial of print developer.
- Remove contents and empty activator packet into container.
- Remove clear tab covering hole on bottom of HotShot™ canister. Place canister into container with hole side down. DO NOT TOUCH THE DRUM. The drum will become very hot. The plastic container will also become very hot and should not be touched either.
- Empty print developer onto top of metal cap of canister or equivalent of about 15 drops. Fumes will begin as the HotShot™ canister heats up. Make certain this is done in a well-ventilated area or fuming chamber.
- After fuming is complete, allow the container to cool down and dispose of contents in an appropriate manner.
- Re-establish air flow and evacuate fumes prior to entering an area that has been processed with cyanoacrylate.

CA Pad or Pack

Use of commercially available cyanoacrylate packs (e.g., Hard Evidence) is slower than heating or chemically accelerated fuming.
development but is easy to use without the necessity for handling chemicals. This is a good choice when the area to be fumed is exceptionally large and time is not a factor.

- Place the opened cyanoacrylate pack in the enclosed area. If the pack is dried out, it should not be used and should be thrown away.
- The addition of humidity to the fuming chamber/enclosed area (such as a vehicle interior) plays a major role in successful development of white ridge detail in latent prints. This may be accomplished by placing warm water in a cup or similar vessel in the fuming chamber/enclosed area.
- Close the fuming ‘chamber’. Monitor the test print at 5 to 10-minute intervals and stop when the test print becomes visible.
- Sometimes over-development will occur. Over-development is usually in the form of a heavy white deposit obscuring most of a latent print. Use of an adhesive lifting technique (e.g., tape, lifter, etc.) is effective in lifting away the heavy upper deposits revealing underlying ridge detail.
- Evacuate the chamber for a period of 10 minutes prior to additional evaluation/processing.

**Fuming Wands**

The fume wand (sometimes called a glue wand) is a manufactured disposable cyanoacrylate ester device.

**Fueling:** Refer to the manual in the kit for specific manufacturer’s instructions.

**Ignition:** Refer to the manual in the kit for specific manufacturer’s instructions.

At the normal (maximum) setting, a dense stream of cyanoacrylate vapors will begin to emerge within a minute. Place the wand through an opening into the target area. For a vehicle interior, lower one window slightly, taping around the window edges until only a small area remains (regular lift tape works well).

Sometimes in vehicles it is necessary to use two or more cartridges. When one cartridge is exhausted, remove the wand from the window and install another cartridge using metal tweezers. Repeat until you believe adequate vapor is present. Once the vapors have filled the area, remove the wand and seal the opening. Allow the vapor to remain in the vehicle or fuming area for at least 15 minutes or until the test print demonstrates sufficient development.

Open the vehicle doors to evacuate all fumes prior to entering the vehicle.
Cautions/Safety
Cyanoacrylate ester fumes are strongly irritating to the eyes and respiratory system. Fuming should only be conducted in a well-ventilated area and non-porous gloves should be worn to prevent skin contact.

Wands:
The fuming wand and cartridges become very hot and should be handled with caution. In some cases, it is difficult to determine if the wand is lit. The flame may not be visible. Thus, one should not rely on whether or not the flame is visible to determine if the wand is still burning.

HotShot™ (or similar) fingerprint developer:
Upon activation, the canister will become very hot. Caution should be used when placing on a heat sensitive surface like plastic. This kit must be used in a well-ventilated area and care must be taken to avoid breathing in vapors.

Latent print development with super glue fumes can be accomplished by creating a fuming chamber (e.g., vehicle interior, improvised enclosed area, plastic bag, fish tank). It should be noted that the cyanoacrylate esters can cause a glaze-like coating to cover the entire evidentiary surface resulting in considerable loss of contrast when over-fuming occurs.

Do not refill butane-fueled wands near a heat source.
Do not store cyanoacrylate in areas that can become hot (e.g., the trunk of a car); the cartridges may start to fume, and the pads/packs or liquid may dry out.

Cyanoacrylate should be allowed to come to room temperature prior to use.

Controls
A positive/negative control will be performed with each use. Controls are conducted at the time of use.

Positive control: place asebaceous print onto a clean black lift card or substrate of choice. The test card is placed within the confines of the area to be fumed (for example, a vehicle interior).

The test card must be visible so that latent print development may be monitored to avoid over-fuming.

Negative control: The area surrounding the intentionally placed print for the positive control.
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**Documentation**
The results, for both the casework and the positive and negative control, shall be documented in the case notes.

**Results and Interpretations**
Any areas of ridge detail that are suitable for collection after fuming should be photographed prior to additional processing with powder.

**7.6.6.4. Small Particle Reagent**
Small particle reagent is used to develop latent prints from a variety of surfaces including adhesives and non-porous items that are or have been wet. The color of SPR should be chosen to contrast with the background.

**Reagent**
- Commercially available SPR
  - Or
  - **Working Solution**
    - Molybdenum Disulfide (MoS2) 30 grams
    - Photo Flo 3-4 drops
    - Distilled water 1000 mL

  Dissolve 30 grams of MoS2 in 1000 mL of distilled water and place on magnetic stirrer. Add Photo Flo to the solution.

**Method**
- Apply the Reagent by dipping or spraying.
- Allow to set for 2-3 minutes.
- Gently rinse with water.
- Allow to air dry.
- Repeat procedure if necessary.
- Evaluate item for comparable ridge detail.
- Prints deemed to be of value are marked, photographed, and can be lifted.

**Cautions/Safety**
- There does not appear to be any health hazards associated with small particle reagent.
- Gloves and safety glasses should be worn.
7.6.6.5. **Elimination Fingerprints**

When fingerprints are found at the scene of the crime, consideration should be given to the possibility that the impressions could belong to someone whose presence is legitimate. Rolled inked impressions (elimination fingerprints) should be taken from members of the household, witnesses, officers, or anyone who might have touched something at the scene.

**Equipment**
- Ink source (i.e., inkpad or ink strips)
- Powder
- White adhesive lifts and clear backings
- Elimination fingerprint and palm print cards
- Powder and lift cards may also be used

**Method**

**Inkpad method**
- Instruct the subject to relax the hand and fingers and let operator do the work.
- Roll one finger on a fresh inkpad placing the finger so that it is inked from below the first joint to a point near the tip of the finger, and from nail edge to nail edge.
- The areas should be covered with a light layer of ink. Lack of ink or excessive ink will yield insufficient prints.
- Roll the finger from nail bed to nail bed in the appropriate space on the elimination card.
- One hand of the operator should be used to grip the subject’s finger between the first and second joint. The operator’s other hand should control the pressure and guide the movement of the finger that is being rolled.
- The roll should occur in one steady movement, using the same amount of pressure.
- The finger is then lifted from the card. Do not ‘rock’ a finger over a print that has already been transferred on the card.
  - Repeat this process for each finger.

**Powder method**
- Powder may also be used – powder the ridge detail and lift with tape/adhesive lift.
- Place lift on a paper or place clear backing on white adhesive lift.
GOOD FINGERPRINT STANDARD

- Fingers rolled nail edge to nail edge
- Information below first joint is recorded on most prints
- Slap prints include part of second joint
- Thumb slaps have been rolled up to record the thumb print (this example can be seen demonstrated mostly on right thumb above)
7.6.6.6. **Elimination Palmprints**
For elimination palm prints collection at crime scenes you have the option of using ink or powder for collection. It is important to ensure that the entire palm of the subject is covered with ink/powder.

**Equipment**
- Ink source (i.e., inkpad or ink strips)
- Powder
- White adhesive lifts and clear backings
- Elimination fingerprint and palm print cards
- Powder and lift cards may also be used

**Method**

**Inkpad method**
- Rolling method: Take a cylindrical tube with the palm print card affixed at an angle: Place the card/tube on a flat surface and place the base of the subject’s inked palm on the base of the card/tube. Roll the card/tube by applying slight pressure to the subject’s palm, allowing the entire palm surface (to the fingertips) to make contact with the card/tube while simultaneously rolling the cylinder backwards.

- Solid surface: Place the subject’s inked palm flat onto the palm print card applying enough pressure to force the surface of palm to make uniform contact with the card. Do not apply so much pressure that the ridges become distorted and detail is lost. Then lift the palm up and allow the ink to dry.

- Fill out all appropriate information on the elimination print cards and make sure the clarity of the prints is sufficient for comparison.

**Powder method Palm**
- Powder may also be used – powder the ridge detail (hand) and lift with tape/adhesive lift.

- Place lift on a paper or place clear backing on white adhesive lift.
7.6.7. Living/Deceased Human Evidence or Standards

Nearly all procedures utilized by the forensic scientist may need to be employed when processing human remains or living humans depending on the type(s) of evidence encountered. When documenting human remains at a crime scene it's important to use proper camera lighting and scales to document injuries on the body.

**Equipment**

- Camera
- Scales
- Evidence bags
- Swabs

**Living/Deceased Human Evidence or Standards**

- Document the position and location of the body within the scene through notes, photographs, sketches and/or measurements to reference points.
- Document transient evidence located on the body (i.e., trace evidence, biological fluid stains/patterns, impression evidence). It is best practice to collect transient evidence prior to the removal of the body. Removal of clothing articles may be necessary to preserve pattern evidence; however, it is highly recommended that the forensic scientist communicate with the Coroner’s Office prior to removing the clothing.
- Careful consideration shall be made for the possible presence of biological evidence that could be left by a perpetrator. Bindings, ligatures, the neck, breasts, ankles, arms, bite-marks, or certain areas of the clothing could contain evidence from strangulation, sexual assault, dragging, pocket-rifling or other types of aggressive contact.
- If swabs are collected from the body or bindings, that information should be disseminated to the investigator and coroner’s office, so that collection is not duplicated at a later time.
- Bindings and ligatures shall not be disturbed unless they attach the body to the scene. Minimize the number of cuts to bindings necessary to release the body from the scene. Never cut through a knot. Tape and label end of ligatures or bindings that were cut by investigators.
- Full inked fingerprint standards should not be collected at the scene. Upon approval from the coroner’s office, one or two inked impressions may be taken at the crime scene for rush identification.
- If there is potential evidence on the hands, place a paper bag over each hand labeling the bags to designate right and left and secure with tape prior to removal of the body.
- The presence of liver mortis, rigor mortis or other decomposition characteristics should be documented when noted. Outdoor and indoor temperatures should be noted in addition to the location and condition of fans, air conditioners, and heaters when observed to be relevant.
- Before the body is removed from the scene, ensure evidence will not be
destroyed along the path of transport.

- If multiple bodies are present at a scene, it is necessary to identify which body specific evidence was collected from with a unique identifier (i.e., name, number).

**Victim, Suspect, and/or Witness Evidence/standards**

**Biological and Trace Evidence**
- Photograph and collect visible trace evidence (e.g., glass, hair) on the person or clothing.
- Examine the hands of the individual for biological transfer evidence (e.g., blood).
- Document and collect any evidence located.

**Clothing Evidence**
- Photograph the clothed individual.
- Take close-up photographs of any pattern or transfer evidence visible on the clothing with and without a scale.
- Document each item of clothing collected and package separately.
- If pattern evidence is an issue, the forensic scientist should be cautious not to fold the clothing onto itself to avoid contamination from one area of the clothing item to another, butcher paper can help with this type of packaging.
- If the evidence is wet, the clothing should be dried before packaging. If this is not practical, package the wet clothing in paper bags.

**Injuries**
- All injuries should be documented through notes, photographs, and measurements as to type of injury (i.e., gunshot wound, laceration, abrasion, contusion, etc.), shape, color, size, and location
- Overall photographs shall be taken of the individual for identification purposes.
- Close-up photographs should be taken of the injury with and without a scale.
- A swab should be collected from recent bite mark injuries for possible DNA analysis.

**Sexual Assault Victims**
Certified medical personnel conduct sexual assault examinations, including collection of a Sexual Assault Forensic Evidence Kit and the victim’s clothing.

**DNA Standard Evidence**
Collect two oral (buccal) swabs by rubbing each swab thoroughly on the inner facial cheek of the individual. Package all (two) swabs together in an envelope and label with the individual’s name.

**Shoeprint Standard Evidence**
Some scenes where shoeprint evidence has been collected will require the collection of elimination shoeprints from victims, suspects, witnesses, and other individuals associated with the scene. Collection of the pair of shoes is preferred, but not always reasonable, especially with witnesses or Emergency Responders. There are several methods used to collect elimination shoeprint impressions.
• Dust the soles of the shoes with fingerprint powder and press the sole onto a sheet of adhesive paper. Place a clear sheet of plastic over the print to protect it.
• Lightly smear the outsoles of the shoes with petroleum jelly then press the outsoles onto a sheet of clean white paper. The impressions may be enhanced by lightly dusting with dark fingerprint powder.
• An inkless shoe print kit can be used by having the individual step onto the inkless pad and then onto the chemical sensitive paper.
• Other products available for use are gel lifters and Biofoam.
• Label the impression(s).

Fingernail Evidence
When a forensic scientist collects samples from the suspect and/or from a victim, if length of the nails permits, the nails should be clipped and collected. Clippings and/or swabs from each hand shall be collected and packaged separately. A moistened swab may be utilized for collection if it is not possible to take cutting or may be used additionally after/in addition to the cuttings. This can be routine or especially if there is visible staining.

Skeletal Recovery
The successful recognition and recovery of human skeletal remains is important in determining the identity of the individual(s), as well as providing investigators with forensic evidence in which to further their investigation. Proper techniques must be employed in order to retrieve as many bones of the human body as possible; this increases the probability of positive identifications based on anomalous features unique to an individual. Forensic scientists shall work in conjunction with the Coroner’s office.

Equipment
• Rakes
• Square and spade shovels
• Plastic trowels
• Paint brushes
• Dental tools
• Tongue depressors
• Survey flags
• Survey paint
• Measuring tape
• Wooden stakes
• Colored string
• Canvas or plastic tarps
• Sifting screens
• Camera
• Grid paper
• Notebook
Method

Scattered Remains
• Document location of remains to include measurements. (a grid system is a preferred system of documentation of evidence in this type of system, specially if a dig is going to be following the recovery of the remains, see below).
• Document condition to include vegetation growth or animal interaction.
• Once remains are removed, examine the area directly underneath remains. The forensic scientist should sift the dirt directly underneath if there is a possibility of evidence being present. X-rays can also be used, if available.

Buried Remains
• Document the environment of the burial (e.g., compact, disturbed, shallow).
• Stake (grid) out a work area around the burial area for diagramming and measuring purposes.
• Locate a fixed point in the landscape to perform the appropriate measurement technique(s). If there is a grid point, select a datum point from this grid point.
• Locate the grave outline by visualizing cracked soil or depressions. Work horizontally.
• Sift the soil level by level.
• Orient the shovel blade so that the ground is scraped away horizontally.
• Document all changes in soil density to include color or texture.
• Stop and document as each item of evidence is discovered.
• Evidence shall not be removed unless it is transitory.
• Stop when the skeletal remains are first discovered.
• Remove enough dirt to determine the orientation of the body.
• Pedestal the skeletal features, i.e., remove all soil around the skeletal elements without disturbing their position.
• Expose without removal and document. Expose all the remains and all the associated evidence.

Autopsy Evidence

The autopsy is very individualized; each case dictates what evidence should be collected and what observations are essential. The Coroner’s Office will determine which examinations are to be conducted at the autopsy; the forensic scientist’s function is to assist with the recognition, documentation, and collection of evidence. The forensic scientist may be asked by the investigator to photograph during the autopsy.

Deceased Prints
• Exemplars shall be taken of all homicide victims unless the decedent’s condition does not allow for collection.
• Where the body may be decomposed, burned, mummified, or in a state of extreme rigor, an appropriately trained Latent Print forensic scientist should be called.
• Refer to the Latent Print analytical method regarding advanced techniques for deceased printing.
7.6.8. Fire Debris

Fire scenes often have extensive of damage to evidence caused by the fire, firefighter efforts, and other personnel. It is important for the forensic scientist to identify who may have altered the scene and in which fire scene this can be complex. Forensic scientists may have the assistance of the State Fire Marshal. In situations where there is no fire marshal to assist, a forensic scientist needs to make sure to look for accelerants, volatiles, combustible, electrical setups, etc.

Package non-liquid potential volatiles and inhalants into leak-proof vapor-tight containers as soon as possible (e.g., metal can or vapor-tight/ “fire debris” bag). Leave a little room (i.e., no more than 2/3 full) and do not pack down the contents. Freezer storage is recommended.

Volatile stes include pepper spray, bear spray, perfumes, or any other sample(s) where the “odor” is coming from the chemical of interest. Reference samples of pepper or bear sprays are not needed for analysis. Controls should be collected from areas were volatiles are suspected of not being placed and packed in the same manner. Cuttings shall be made with clean blades between samples to limit contamination.

7.7. Evaluation of Measurement of Uncertainty

This does not apply to crime scene processing.

7.8. Ensuring the Validity of Results

7.8.1. Procedure, controls, and standards for monitoring tests are included in each method. Results are recorded in the case notes and/or quality records (e.g., Reagent log), as appropriate.

7.8.2. See the ACSO Quality Manual for review criteria.

7.9. Reporting the Results

7.9.1. Measurements reported and collected during processing of crime scene processing (scene measurements, trajectories, bloodstain, etc..) are used as descriptors and are not meant to be interpreted as quantitative forensic test results. Any reported measurement is approximate.

7.9.2. Results reported relate only to the items tested. Additionally, a reporting and review guide may be utilized to ensure appropriate notes and report content.
Ada County Sheriff’s Office Forensic Lab

7.9.3. Report is to include all tests performed, tools utilized, and items collected

7.9.4. Final paragraph statement in the field service report:

7.9.4.1. All photographs and collected evidence were/was properly secured and transported to the Ada County Sheriff’s Office. Evidence requiring forensic laboratory analysis will be submitted to the appropriate forensic laboratory upon request.

7.9.5. Notes to add if applicable in field service reports:

7.9.5.1. For the purpose of this report, any blood reported is based on visual examination only and has not been confirmatory or presumptively tested unless otherwise noted.

7.9.5.2. Any fingerprints developed as a part of crime scene processing must be further analyzed for suitability to compare. Prints collected may not be of value once further latent fingerprint examination is conducted.

7.10. **Complaints**

See ACSO Forensic Lab Quality Assurance Manual

7.11. **Non-conforming Work**

See ACSO Forensic Lab Quality Assurance Manual

7.12. **Control of Data and Information Management**

See ACSO Forensic Lab Quality Assurance Manual

8.0 **Management System Requirements**

See ACSO Forensic Lab Quality Assurance Manual
## History of Crime Scene Analytical Method

<table>
<thead>
<tr>
<th>Section &amp; Comments</th>
<th>Date Adopted</th>
<th>Author</th>
<th>Reviewer(s)</th>
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<td>7.13.2018</td>
<td>NW</td>
<td>EH, KB, LK, and HC</td>
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<td>Document all requested work to be performed. Document any deviations from that request and communication regarding the deviation.</td>
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<td>4.13.7.7.7 added Or annotation</td>
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<td>5.4.2.7.3 added inconclusive conclusion</td>
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<td>5.4.5.2 clothing should be collected instead of shall</td>
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<td>5.4.7.5.4 Close ups should be taken instead of shall</td>
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<td>Re-write of manual formatted to ISO/IEC 17025:2017 standards and ANAB 3125 accreditation requirements and numbering format. Photography quality and procedures were updated, cameras were considered not equipment needed testing.</td>
<td>4.19.2021</td>
<td>EH</td>
<td>NW, KB, LK, HC, and SG</td>
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<td>Edited metal detector information.</td>
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<td>Large scale changes and added modification throughout the entire Analytical Method.</td>
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